Cool-temperature-mediated activation of phospholipase C-[gamma]2 in the human hereditary disease PLAID

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Citation: SCHADE, A. ... et al., 2016. Cool-temperature-mediated activation of phospholipase C-[gamma]2 in the human hereditary disease PLAID. Cellular Signalling, 28 (9), pp. 1237–1251.

Additional Information:

- This paper was accepted for publication in the journal Cellular Signalling and the definitive published version is available at http://dx.doi.org/10.1016/j.cellsig.2016.05.010.

Metadata Record: https://dspace.lboro.ac.uk/2134/21459

Version: Accepted for publication

Publisher: © Elsevier

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Dear Professor Maurice,

thank you very much for the evaluation of and very positive response to our manuscript "Cool-temperature-mediated activation of phospholipase C-γ2 in the human hereditary disease PLAID" by three expert reviewers of Cellular Signalling and for allowing us to submit a revised version of our work to the Journal. Thank you also very much for the extension of the deadline (hoping that you will allow me to use Canadian time and date for the submission).

In the following, we provide a point-by-point reply to the critical points raised by the reviewers:

(i) Reviewer 1: "The results section of the manuscript lacks subtitles. Addition of these would enhance the stringency of the presentation of the results and help the reader to digest the data more easily."

Authors’ response: This was done in the revised manuscript.

(ii) Reviewer 1: "In all the figures where temperature shifts were analyzed, the authors choose the conventional way, where left starts with the lower temperature and right on the x axis are the higher temperature values. However, my personal opinion is that it would be more intuitive for the reader to put it the other way round. Left the physiologic temperature and right the reduced temperatures, which than lead to enhanced mutant signals. At least for me this would be a more intuitive way to show the data."

Authors’ response: Although we clearly see and understand the point of the reviewer, we would very much like to adhere to the previous mode of presentation. This is mostly due to the fact that we feel that gradual changes in the temperature, such as those used in many of the figures (rather than the step changes used in the other figures) are clearer in the low to high temperature left to right presentation commonly used in physics. In particular, this format is commonly used for Q10 presentations such as the one in Fig. 3 in the field of temperature regulation of other proteins, e.g. of TRP channels.

(ii) Reviewer 1: "At least in the discussion the authors should address the question of temperature vs. ligand induced activation of Plcg2. The experiment with the constitutive Rac as activator is not entirely convincing. In vivo in mutant mice and in the patient the disease phenotype might be a mixture of temperature and ligand induce overreactivity, as the different mutants suggest." This point is related to Minor Point of Reviewer 2: "It would be interesting to know if upon stimulation (BCR triggering) at low temperature the highly active mutants can be further stimulated."
Authors’ response: We have followed these important suggestions as much as we could possibly do in the COS-7 cell context, by analyzing the activation of exogenous wild-type versus Δ20-22 and Δ19 mutant PLCγ2 by EGF receptors endogenously present in these cells. The results, which are shown in Fig. 11 of the revised manuscript, are intriguing because they imply that the mutant PLCγ2 enzymes are resistant to stimulation by EGFR activation, both at 31 °C and 37 °C. This is behavior is in striking contrast to that of wild-type PLCγ2 with approx. 12-fold stimulation by EGF receptor activation at both 31 °C and 37 °C. Although activation of PLCγ2 in COS-7 by activated EGFR differs from its activation in B lymphocytes by antigen-activated BCR or in other cells by other receptor tyrosine kinases, these results are similar to the loss of function effects seen in cells of PLAID patients and could provide hints to the mechanistic basis of the apparently disparate effects of the PLAID mutations, gain and loss of PLCγ2 function.

(iii) Reviewer 1: "The octamer peptide PCI is fully conserved in PLC<gamma>1 and mediates cold sensitivity in PLC<gamma>2. Which effect causes its deletion in PLC<gamma>1."

Authors’ response: We have not specifically addressed this question. However, since the PCI peptide is contained in the region deleted in the PLCγ1 mutant "Δ20-22", we would predict that the functions of PLCγ1Δ PCI would be similar to those of PLCγ1Δ20-22 (cf. Fig. 4A), such as the functions of PLCγ2Δ PCI resemble those of PLCγ2Δ20-22 (cf. Fig. 4B, left). However, we do not feel that the octamer mediates cold sensitivity in PLCγ2, since it is maintained in PLCγ2Δ19, which is nevertheless sensitive to activation by cooling (cf. Fig. 8B).

(iv) Reviewer 1: "The authors could provide data on the mobilization of Ca2+ upon the temperature shift. This would strengthen the data on IP3 measurements."

Authors’ response: We appreciate this suggestion of the reviewer. However, in other studies that we have performed on PLCγ2, we have always seen a close correlation between inositol phosphate formation and increase in cytosolic Ca2+, at least upon acute activation of the enzyme. In the chronic activation setting used in most of the experiments shown here, we would anticipate technical difficulties in observing reliable changes in [Ca2+]i for the PLCγ2 PLAID mutants.

(v) Reviewer 3: "Attempts were made to provide a structural rational for the activation mediated by cool temperatures. The main limitation here is that the work relies on indirect experiments that measure enzyme activity of PLCgamma2 variants in transfected cells without any further support from other approaches. Conceptual schemes are helpful but experimental limitations have to be pointed out. This should be emphasized in discussion."

Authors’ response: This was explicitly done on p. 19 of the revised manuscript.

(vi) Reviewer 3: "Several observations deserve some more emphasis even if they can not be easily explained at present. These include: -The data suggesting that PLCgamma1 variants harboring deletions are not regulated by cool temperatures and that regions corresponding to deletions in PLAID have a bigger role in auto-inhibition in this enzyme. -The data related to Ali5 mutation and activation by cool temperatures. This mutation is not in the region affected by PLAID, actually not in the regulatory region at all. Some link to the PH domain has been suggested but this is mechanistically unclear. Unexpected behavior of Ali5 variant on its own and the distinct position of the mutation need to be pointed out. -The fact that many deletions in PLCgamma2 SH region are further activated by cool temperatures."

Authors’ response: All three points have been addressed in the text of the revised manuscript on pp. 11, 13 (top paragraph), and 13 (bottom paragraph).

(vii) Reviewer 3: "It should be stated that assumption (or expectation) has been made that observations based on COS cell transfections will be relevant for endogenous expression levels in B-cells."
Authors’ response: This was done in the revised manuscript on p. 20 in the context of discussing the resistance of the two deletion mutants to activated EGFR in COS-7 cells [cf. (ii)].

An additional point not requested by the reviewers was the examination of the requirement of the spPH domain per se for cool temperature regulation of PLCγ2. We felt that this experiment needed to be done and think that its results should be reported in this manuscript (now in Fig. 10B) to avoid misinterpretations of the role of spPH in mediating the enzyme’s response to cooling. The results are intriguing: while spPH exerts a striking regulatory role within the PLAID PLCγ2 mutant PLCγ2Δ19, it is not required for the cool temperature response of further truncated mutants, such as the bipartite mutant made up of fragments X and Y. We would be happy, if you allowed us to show this data and an accordingly revised version of the model now shown in Fig. 10C-H.

In summary, we believe that we have carefully considered and taken care of most, if not all of the referees’ critical points. We sincerely hope that these changes make our work acceptable for publication in Cellular Signalling. Needless to say, we are extremely grateful to all three reviewers for their expert opinions and invaluable and important advice on the manuscript.

Yours sincerely,

Peter Gierschik
Cool-temperature-mediated activation of phospholipase C-γ2
in the human hereditary disease PLAID

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Abbreviations: PLC, inositol-phospholipid-specific phospholipase C; SH2, Src homology domain 2; SH3, Src homology domain 3; PH, pleckstrin homology domain; spPH, split PH domain; PCI, phospholipase C inhibitor peptide; Rac, Ras-related C3 botulinum toxin substrate; BCR, B cell receptor; SA, specific array; PLAID, PLCγ2-associated antibody deficiency and immune dysregulation; TRP, transient receptor potential; PtdIns, phosphatidylinositol; PtdIns(4)P, phosphatidylinositol 4-phosphate; PtdIns(4,5)P2, phosphatidylinositol 4,5-bisphosphate; Co., control; aa, amino acid

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**ABSTRACT**

Deletions in the gene encoding signal-transducing inositol phospholipid-specific phospholipase C-γ2 (PLCγ2) are associated with the novel human hereditary disease PLAID (PLCγ2-associated antibody deficiency and immune dysregulation). PLAID is characterized by a rather puzzling concurrence of augmented and diminished functions of the immune system, such as cold urticaria triggered by only minimal decreases in temperature, autoimmunity, and immunodeficiency.

Understanding of the functional effects of the genomic alterations at the level of the affected enzyme, PLCγ2, is currently lacking. PLCγ2 is critically involved in coupling various cell surface receptors to regulation of important functions of immune cells such as mast cells, B cells, monocytes/macrophages, and neutrophils. PLCγ2 is unique by carrying three Src (SH) and one split pleckstrin homology domain (spPH) between the two catalytic subdomains (spPHn-SH2n-SH2c-SH3-spPHc). Prevailing evidence suggests that activation of PLCγ2 is primarily due to loss of SH-region-mediated autoinhibition and/or enhanced plasma membrane translocation. Here, we show that the two PLAID PLCγ2 mutants lacking portions of the SH region are strongly (> 100-fold), rapidly, and reversibly activated by cooling by only a few degrees. We found that the mechanism(s) underlying PLCγ2 PLAID mutant activation by cool temperatures is distinct from a mere loss of SH-region-mediated autoinhibition and dependent on both the integrity and the pliability of the spPH domain. The results suggest a new mechanism of PLCγ activation with unique thermodynamic features and assign a novel regulatory role to its spPH domain. Involvement of this mechanism in other human disease states associated with cooling such as exertional asthma and certain acute coronary events appears an intriguing possibility.

**Keywords:** Phospholipase C-γ2; Inositol phospholipid; Rac2 GTPase; Split PH domain; Autoinhibition; Cold temperature sensitivity
1. Introduction

Inositol-phospholipid-specific phospholipases C (PLCs) catalyse the formation of inositol 1,4,5-trisphosphate and diacylglycerol, and, at the same time, decrease the local or general plasma membrane abundance of their substrate, phosphatidylinositol 4,5-bisphosphate (PtdIns$_{P2}$) [1]. The latter three molecules are important mediators of cellular signaling. An enormous variety of cell surface receptors regulates important cellular functions utilizing PLCs, ranging from G-protein-coupled receptors over certain ion channels to many transmembrane non-enzymes and enzymes, e.g. receptor tyrosine kinases. The mammalian PLCs are divided into six subfamilies, β, γ, δ, ε, ζ, and η. Analyses of PLC crystal structures have revealed that the catalytic mechanism of PLCs is well conserved between all PLC family members. Their activation by cell surface receptors involves both translocation of the soluble PLC enzymes to the plasma membrane, i.e. the site of their phospholipid substrate(s), and removal of intramolecular autoinhibition [2-5]. However, certain findings suggest further, still unknown regulatory mechanisms of PLC isozyme activation [1].

The two members of the PLCγ subfamily, PLCγ$_1$ and PLCγ$_2$, control functions represented in many, if not all cell types, such as cell growth and differentiation, as well as migratory responses, but are also involved in highly specialized tasks [6]. Examples of the latter are those regulated by PLCγ$_2$ in cells of the immune system. PLCγ$_1$ and PLCγ$_2$ are activated by receptor and nonreceptor tyrosine kinases; PLCγ$_2$ is also activated by Rac GTPases [7]. In B lymphocytes, this interaction amplifies B-cell-receptor-mediated Ca$^{2+}$ signalling [8]. The structures of the two PLCγ isozymes are unique in that the two catalytic subdomains X and Y are separated by a modular assembly comprising a split PH domain (spPH$_n$ and spPH$_c$), two SH2 (SH2$_n$ and SH2$_c$), and one SH3 domain. The whole assembly (spPH$_n$-SH2$_n$-SH2$_c$-SH3-spPH$_c$) is also referred to as specific array (γSA). Studies on isolated γSA structures showed that the split PH domains of PLCγ$_1$ and PLCγ$_2$ do not alter their three-dimensional shapes upon insertion of the entire PLCγ$_1$ SH2$n$-SH2$c$-SH3 region between the two PLCγ$_1$ spPH halves and upon peptide ligand binding to the insertion [9] or, in case of spPH of PLCγ$_2$, upon its interaction with activated Rac2 [10]. These findings suggested that the PLCγ split PH domain is a more rigid, conformationally stiff element of γSA and that it mediates PLCγ$_2$ activation mainly by allowing Rac2 to translocate the enzyme to the plasma membrane. Recent evidence suggests that the SH2$c$ domain is a major determinant of PLCγ$_1$ autoinhibition and that activation of the enzyme by tyrosine phosphorylation at a site immediately downstream of the domain (Y$_{783}$) proceeds by competition of the phosphorylated peptide with a so far unidentified site on the catalytic XY TIM barrel for binding to SH2$c$ [11,12].
Alterations of the primary structures of PLCγ1 and PLCγ2 are involved in disease, both in humans and in animal models. Thus, point mutations in the human PLCG1 gene have been linked to secondary, radiation-associated angiosarcoma [13] and to cutaneous T cell lymphoma [14]. Two mouse models of autoimmunity and autoinflammation, designated Ali5 and Ali14, have been described, which are caused by gain-of-function point mutations of PLCγ2, D993G and Y495C, respectively [15-17]. The Ali5 mutation also gives rise to platelet hyperreactivity and a prothrombotic phenotype in mice [18]. Recently, deletion of exon 19 or exons 20-22 of the human PLCG2 gene has been shown to cause a novel human hereditary disease characterized by cold urticaria, immunodeficiency, and autoimmunity, designated PLAID for PLCγ2-associated antibody deficiency and immune dysregulation [19]. In affected individuals, only very subtle skin cooling, such as the one caused by a single tear rolling down the cheek at room temperature, causes urticarial wheals and flares within one minute [20]. A related, but distinct human disease, predominantly characterized by autoinflammation and designated APLAID for autoinflammatory PLAID, is caused by a gain-of-function point mutation, S707Y, located in SH2c of PLCγ2 ([21], cf. [22], for a more comprehensive review). Although some effects of decreasing temperature on functions downstream of PLCγ2 have been documented in cells from PLAID patients [19], understanding of cool temperature regulation of the enzyme that is affected by the deletion mutations at first hand, PLCγ2, is currently lacking. Here, we show that the two PLCγ2 mutants identified in PLAID patients, PLCγ2Δ19 and PLCγ2Δ20-22 (Fig. 1A), are exquisitely sensitive to cooling and that the magnitude of the response is unprecedented in that it goes far beyond those previously observed for many other signaling proteins sensitive to temperature changes, such as the transient receptor potential (TRP) cation channels [23]. The results suggest that PLAID PLCγ2 mutants are activated by only minute decreases in temperature by a novel mechanism that is primarily mediated by the split PH domain and distinct from a loss of autoinhibition.

2. Material and methods

2.1. Material

The mouse monoclonal antibody 9B11 reactive against the c-Myc epitope (EQKLISEEDL) and the polyclonal antiserum reactive against human PLCγ2 raised in rabbits (sc-407) were obtained from Cell Signaling Technology and Santa Cruz, respectively. The mouse monoclonal antibody AC-15 reactive against β-actin (A1978), human epidermal growth factor (E9644), and cycloheximide (C7698) were obtained from Sigma.
2.2. Construction of vectors

The construction of complementary DNAs encoding c-Myc-epitope-tagged human PLCγ1 (1291 aa, accession number ABB84466), human PLCγ2 (1265 aa, accession number NP_002652), and the spPH domain chimera PLCγ2-PH1 is described in [24]. The cDNAs of PLCγ2Δ19 (deletion of exon 19, aa 646-685), PLCγ2Δ19-PH12, PLCγ2Δ19-PH21, PLCγ2Δ19-PH11 [with one or both portions of the PLCγ2 spPH domain (aa 468-513 and aa 849-914, respectively) replaced in PLCγ2Δ19 by the corresponding regions of PLCγ1 (aa 482-527 and aa 872-937, respectively)], PLCγ1Δ19 [deletion of PLCγ1 residues corresponding to residues 646 to 685 of PLCγ2 (aa 668-707)], PLCγ2ΔSH (deletion of the SH2n-SH2c-SH3 region, aa 515-840), PLCγ2ΔSH2c (deletion of the C-terminal SH2 domain, aa 639-766), and PLCγ2ΔPCI (aa 726-733) were constructed by *in vitro* mutagenesis using the QuikChange II XL Site-Directed Mutagenesis Kit (200521, Agilent Technologies). The deletion of exons 20-22 in PLCγ2 (PLCγ2Δ20-22, aa 686-806), of the PLCγ1 residues corresponding to residues 686 to 806 of PLCγ2 (aa 708-828; PLCγ1Δ20-22), and of the PLCγ2 specific array (PLCγ2ASA, aa 476-908) was performed using the PCR overlap extension method. The introduction of point mutations was performed by *in vitro* mutagenesis using the QuikChange II XL Site-Directed Mutagenesis Kit according to the manufacturer’s instructions. For the insertion of γ2 SH domains into PLCγ2ΔSH, a linker containing an AvrII restriction site (GGCCCTAGG, AvrII site underlined) was introduced into the deletion site of PLCγ2ΔSH. The SH domains (SH2n, aa 515-638, SH2n, aa 637-747, SH3, aa 767-839, SH2n-SH2c, aa 515-747, SH2c-SH3, aa 637-840) were amplified with primers containing an AvrII restriction site on either end and inserted into PLCγ2ΔSH by restriction and ligation. The linker introduced five additional residues in positions 515-519 of the protein. There were no functional differences between the PLCγ2ΔSH mutants with and without these residues.

Complementary DNAs encoding PLCγ2-XPHn (aa 1-514) and PLCγ2-YPHC (aa 841-1265) were amplified by PCR. Both fragments were c-Myc-tagged at their C-termini. DNAs encoding PLCγ2-X (aa 1-470) and PLCγ2-Y (aa 914-1265) were amplified by PCR. PLCγ2-X and PLCγ2-Y were c-Myc-tagged at their N- and C-termini, respectively. The primer sequences and PCR protocols are available from the authors upon request.

2.3. Cell culture and transfection

COS-7 cells were maintained at 37 °C in a humidified atmosphere of 90 % air and 10 % CO2 in Dulbecco’s modified Eagle’s medium (41965-039, Gibco) supplemented with 10 % (v/v) fetal calf serum (10270-106, Gibco) and 2 mM glutamine, 100 units/ml penicillin, and 100 µg/ml streptomycin (all from PAA Laboratories, Cölbe, Germany). Prior to transfection, COS-7 cells were seeded into 24-well plates at a density of 0.75 x 10^5 cells/well, and grown for 24 h in 0.5 ml of medium/well. For
transfection, plasmid DNA (500 ng/well) was diluted in 50 µl jetPRIME® buffer and 1 µl of jetPRIME® Reagent (114-15, Polyplus Transfection, Illkirch, France) was added according to the manufacturer’s instructions. The total amount of DNA was maintained constant by adding empty vector. Four h after the addition of the DNA-jetPRIME® complexes to the dishes, the medium was replaced by fresh medium, and the cells were incubated for a further 20 h at 37 °C and 10 % CO₂.

2.4. Radiolabeling of inositol phospholipids and analysis of inositol phosphate formation

Twenty four hours after transfection, COS-7 cells were washed once with 0.3 ml/well of Dulbecco’s PBS (PAA Laboratories) and then supplied with 0.2 ml/well of Dulbecco’s modified Eagle’s medium containing supplements as described under Cell culture and transfection, and additionally supplemented with 25 mM HEPES and 2 mM sodium pyruvate (both from PAA Laboratories) to maintain the pH of the medium [25], 2.5 µCi/ml myo-[2-3H]inositol (NET1156005MC, Perkin-Elmer), and 10 mM LiCl. The cells were incubated for 20 h in this medium in individual incubation chambers in ambient atmosphere at temperatures ranging from 25 °C to 39 °C, washed once with 0.2 ml/well of Dulbecco’s PBS, and then lysed by addition of 0.2 ml/well of 10 mM ice-cold formic acid. After keeping the samples for 30 min at 4 °C, 0.3 ml/well of 10 mM NH₄OH was added for neutralization, and the sample was centrifuged for 5 min at 20,000 x g. The supernatants were loaded onto columns containing 0.5 ml of Dowex® 1 x 8-200 ion exchange resin (217425, Sigma) that had been converted to the formate form and equilibrated with H₂O. The columns were washed once with 3 ml of H₂O and twice with 3.5 ml each of 60 mM sodium formate and 5 mM sodium tetraborate. Inositol phosphates were eluted with 3 ml of 1 M ammonium formate and 100 mM formic acid. The eluate was supplemented with 15 ml of scintillation fluid (Quicksafe A, 1008000, Zinsser Analytic, Frankfurt, Germany) and the radioactivity was quantified by liquid scintillation counting.

2.5. Construction of individual incubation chambers

Chambers allowing the temperature-controlled incubation of individual 24-well tissue culture plates (92424; TPP, Switzerland) were custom assembled using 15 x 12 x 23 cm Styrofoam containers (Schaumaplast, Reilingen, Germany) equipped with circuits made up of one Velleman VM148 thermostat control module (190655-62) and one Dallas DS18S20-55 temperature sensor with digital output (176168-62) to control two serially connected heating foils (532878-62; all from Conrad Electronic, http://www.conrad.de) to be placed on either side of the tissue culture plate during incubation. To allow for additional external, analogous control of the temperature, a Ø 10 mm hole was drilled into the wall of the container and the tissue culture plate to allow insertion of a thermometer into one well. Power supply to up to 8 individual chambers was through a TDK-Lambda
LS75-12 AC/DC converter unit (511823-62; Conrad). A construction guidance including a circuit diagram is available from the authors on request.

2.6. Inositol phospholipid analysis

Inositol phospholipids were extracted from transfected cells and analyzed as before [17]. COS-7 cells were grown and radiolabeled in 6-well plates. At the end of the radiolabeling procedure, 10 µl of the medium supernatant were placed in a scintillation vial with 3 ml of Quicksafe A liquid scintillator (Zinsser Analytic) and the radioactivity was quantified by liquid scintillation counting. The radiolabeled cells were lysed by addition of 1.2 ml of 4.5 % (v/v) perchloric acid. After incubating the samples for 30 min on ice, they were scraped into 1.5 ml reaction tubes and centrifuged at 4 °C for 20 min at 3,700 × g. Supernatants and pellets were separated. The pellets were resuspended in 100 µl of water and 375 µl of chloroform/methanol/HCl (100:200:15) was added. The samples were vortexed, and an additional 125 µl of chloroform and 125 µl of 0.1 M HCl were added. After further vortexing, the samples were centrifuged at room temperature for 10 min at 700 × g. Fifty µl each of the lower, chloroformic phase containing the inositol phospholipids were subjected to liquid scintillation counting as described above.

2.7. Determination of the 10-degree temperature coefficients

According to Hille [26], the 10-degree temperature coefficient, $Q_{10}$, of a biological process can be calculated for an arbitrary temperature interval $\Delta T$ from

$$Q_{\Delta T} = (Q_{10})^{\Delta T / 10}$$

Using $\Delta T = T_i - T_{ref}$ and $Q_{\Delta T} = \frac{A_i}{A_{ref}}$, where $T_i$ and $T_{ref}$ are the individual and a reference temperatures and $A_i$ and $A_{ref}$ are the individual and a reference activity, this equation can be rewritten to

$$\log_{10} \left( \frac{A_i}{A_{ref}} \right) = 0.1 (T_i - T_{ref}) \log_{10}(Q_{10})$$

$$\log_{10} \left( \frac{A_i}{A_{ref}} \right) = 0.1 \log_{10}(Q_{10}) T_i - 0.1 \log_{10}(Q_{10}) T_{ref}$$

Upon plotting the $T_i$ vs. the $\log_{10} \left( \frac{A_i}{A_{ref}} \right)$, the $Q_{10}$ value(s) can be calculated from the slopes of the linear portions of the resultant graphs.
2.8. Miscellaneous

Curve fitting was done using GraphPad Prism, version 4.03 (GraphPad Software, San Diego, CA, USA). In Fig. 3, the global curve fitting procedure was applied, where the extra sum of squares F-test is employed to determine whether the best-fit values of the two parameters differ between data sets. The simpler model was selected unless the P value was less than 0.05. The propagation of errors was calculated as outlined in [27]. The various PLC\(_\gamma\) isoforms were tested in this study multiple times in many different combinations, including many that are not presented in this work. The experiments specifically shown herein were repeated two to three times throughout. Data from representative experiments are presented as means ± standard error of triplicate determinations. Control curves were done in Figs. 4B and 7 to confirm that results shown in different panels of the same figure are comparable. The intensities of immunoreactive bands on Western blots corresponding to PLC\(_\gamma_2\) isozymes carrying a c-Myc epitope were within the range allowing semiquantitative comparisons, as shown by control experiments using purified c-Myc-tagged wild-type PLC\(_\gamma_2\). Samples to be analyzed by Western blotting were taken, quasi as a fourth replicate, from the same plate as and immediately adjacent to the samples taken in triplicate for functional analysis. Using this protocol and paying meticulous attention to experimental detail, we have not experienced variations in gel loading of these samples (cf. Fig. 5B).

3. Results

3.1. The PLC\(_\gamma_2\) deletion mutants PLC\(_\gamma_2\)D19 and PLC\(_\gamma_2\)D20-22 are specifically activated by cool temperatures

PLC\(_\gamma_2\)D19 and PLC\(_\gamma_2\)D20-22 were expressed in COS-7 cells at 37 °C and the cells were then radiolabeled with \(^{1}H\)inositol at temperatures ranging from 39 °C to 25 °C, followed by measurement of inositol phosphate formation. Fig. 1B shows that the two PLC\(_\gamma_2\) deletion mutants caused slight increases in inositol phosphate formation in comparison to the wild-type enzyme at 37 °C. Consistent with earlier results [19], these increases were approximately 2.8- and 3.6-fold for PLC\(_\gamma_2\)D19 and PLC\(_\gamma_2\)D20-22, respectively, relative to the increase over basal activity observed for wild-type PLC\(_\gamma_2\). Much more strikingly, however, there was a marked stimulation of inositol phosphate formation when cells expressing either deletion mutant were incubated at only slightly lower temperatures. In both cases, there was a biphasic stimulatory response with declining temperatures, with a maximum at 31 °C and a gradual reduction upon further cooling to 25 °C. There was only a modest monophasic decrease in inositol phosphate formation in cells expressing wild-type PLC\(_\gamma_2\), and in control cells when the temperature was reduced from 39 °C to 25 °C. Strikingly, at 31 °C, the absolute increase in inositol phosphate formation in cells expressing PLC\(_\gamma_2\)D19 and PLC\(_\gamma_2\)D20-22 over basal activity of mock-
transfected control cells was enhanced approx. 480 ± 91- and 430 ± 84-fold (means ± SEM) relative to the increase over basal inositol phosphate formation observed in cells expressing wild-type PLCγ2.

Fig. 1C shows that the expression of wild-type PLCγ2 decreased with decreasing incubation temperature, while the expression of both PLCγ2Δ19 and PLCγ2Δ20-22 took slight increases at intermediate temperatures ranging from 35 °C to 31 °C and from 35 °C to 25 °C, respectively. While reduced expression of wild-type PLCγ2 at lower temperatures may explain, at least in part, the monophasic decrease in inositol phosphate formation by this enzyme, the limited magnitude of the changes observed in Fig. 1C for PLCγ2Δ19 and PLCγ2Δ20-22 argues against a critical role of fluctuating enzyme expression in the marked changes of inositol phosphate formation enzyme activity evident in Fig. 1B.

3.2. The activation of PLCγ2Δ19 and PLCγ2Δ20-22 by subphysiological temperatures occurs after protein synthesis and is reversible.

The influence of a decrease in incubation temperature on the abundance of inositol phospholipids in mock-transfected COS-7 cells and cells expressing either wild-type or Δ19 mutant PLCγ2 is shown in Figs. 1D and E. PLCγ is capable of hydrolyzing PtdIns, PtdIns(4)P, PtdIns(4,5)P2 [28]. There was no increase in the abundance of PLCγ substrate inositol phospholipids upon cooling. Instead, a monophasic loss was apparent in all three cases upon decreasing the temperature to 25 °C to approximately 13 % of the levels observed at 37 °C. This behavior was largely independent of whether or not wild-type PLCγ2 or PLCγ2Δ19 was expressed in the cells. While this loss may reflect slower synthesis of the substrate phospholipids at lower temperatures and may explain some of the decline in inositol phosphate formation by PLCγ2Δ19 and PLCγ2Δ20-22 at temperatures below 31 °C (cf. Fig. 1B), it argues against increased substrate availability causing the dramatic stimulation of the PLCγ2 deletion mutants upon cooling from 37 °C to 31 °C (cf. Fig. 1B).

The next experiment was designed to examine whether stimulation of PLCγ2Δ19 and PLCγ2Δ20-22 by cool temperatures occurs before or after synthesis of the mutant PLCγ2 proteins. To this end, cells already containing either wild-type PLCγ2, PLCγ2Δ19, or PLCγ2Δ20-22 (as a result of a 24-h-transfection period at 37 °C) were treated for a further 20 h (i.e. during the [3H]inositol radiolabeling phase) with or without cooling from 37 °C or 31 °C, both in the absence and in the presence of 100 μg/ml cycloheximide. Using this protocol, we expected that cycloheximide would block the stimulatory effect of cooling on inositol phospholipid hydrolysis by the mutants, if cooling were to be effective prior to or during recombinant protein synthesis. Fig. 2A shows that this was not the case. Thus, cooling to 31 °C caused marked increases in inositol phosphate formation, regardless of whether cycloheximide was absent or present during the second phase. Fig. 2B shows that, both at 37 °C and
at 31°C, cycloheximide prevented the increase in abundance of wild-type and deletion mutant PLCγ2 during the second phase (in comparison to the control samples obtained after the initial phase), indicating that cycloheximide was in fact effective as a protein biosynthesis inhibitor in this experiment. Taken together, these results indicate that the stimulatory effect of subphysiologic temperatures on deletion mutant PLCγ2 activity occurs after protein synthesis.

To examine whether the stimulatory effects of the Δ19 and Δ20-22 deletions on PLCγ2 activity were reversible, wild-type PLCγ2, PLCγ2Δ19, and PLCγ2Δ20-22 were incubated for the last four hours of the transfection protocol, i.e. in the absence of radiolabeled inositol phospholipid substrate, at either 37°C or 31°C and then radiolabeled with [3H]inositol at one of the two temperatures. Fig. 2D shows that the three different incubation protocols had little, if any, effect on the expression of each of the three enzymes. As shown in Fig. 2C, preincubation of the three enzymes at 31°C had no effect on their ability to promote inositol phosphate formation. Only when cells were incubated at 31°C in the presence of [3H]inositol, enhanced inositol phosphate formation was evident by the two deletion mutants, in contrast to wild-type PLCγ2. A time-course of the changes of inositol phosphate formation by wild-type and mutant PLCγ2 upon changes in incubation temperature from 37°C to 31°C and vice versa is shown in Figs. 2E and 2F. Both deletion mutants were activated with no apparent lag time with cooling to 31°C. However, deactivation of both enzymes by warming to 37°C required a lag time of approximately 10 min to come into effect. Thus, activation of PLCγ2Δ19 and PLCγ2Δ20-22 by cool temperatures is a rapid and slowly, but fully reversible process.

The determination of the 10°C temperature coefficient (Q10) value, widely used to characterize the regulation of TRP channels by temperature [23], for wild-type PLCγ2, PLCγ2Δ19, and PLCγ2Δ20-22 is shown in Fig. 3. While only a single linear component with a Q10 value of 4.6 was evident for wild-type PLCγ2 between 39°C and 25°C, both PLCγ2Δ19 and PLCγ2Δ20-22 displayed two separate phases of opposite signs and markedly distinct Q10 values. Specifically, while Q10 was as high as 6745 for the marked increase in activity between 39°C and 33°C, it was lower by more than three orders of magnitude and not different, by sign and by magnitude, from the Q10 value describing the decrease in activity of the wild-type enzyme, 4.6, between 31°C and 25°C. Thus, it appears that the major functional consequence of the two deletions is their increased activity upon cooling from 37°C to approximately 31°C. The decrease in activity observed at temperatures below 31°C appears to be a property inherently present in the remaining elements of wild-type PLCγ2. The Q10 values of PLCγ2Δ19 and PLCγ2Δ20-22 exceed those reported for other temperature-regulated proteins, such as thermoTRPs (with values ranging from 10 to > 100 [23]). However, higher Q10 values are not without precedence. For example, the step of heat damage to the development of the posterior crossvein of Drosophila that is most sensitive to increasing temperature showed a Q10 of about 360 [29].
3.3. Cool-temperature-mediated activation of PLCγ2D19 and PLCγ2D20-22 is distinct from loss of SH-region-mediated autoinhibition.

The structural organization of PLCγ2 between its two catalytic subdomains X and Y is very similar to that of its close relative PLCγ1 [6]. To address the question, whether deletion in PLCγ1 of residues corresponding to those encoded by exons 19 and 20-22 of PLCG2 has similar functional consequences, the relevant mutants of human PLCγ1 were produced. Since the human PLCG1 and PLCG2 genes exhibit distinct patterns of genomic organization, the deletions in PLCγ2 do not correspond to specific exons in PLCG1. Hence, the mutants are designated PLCγ1Δ19” and PLCγ1Δ20-22”. Fig. 4A shows that both PLCγ1Δ19” and PLCγ1Δ20-22”, in contrast to their PLCγ2 counterparts, were already highly constitutively active at 37 °C (54- and 37-fold increases in inositol phosphate formation, respectively, in comparison to the increase caused by wild-type PLCγ1 relative to control). Furthermore, only minor changes in their activity were observed upon reduction of the incubation temperature from 37 °C to 27 °C. There were only minor changes in inositol phosphate formation by wild-type PLCγ1. Fig. S1A shows that there were minor, if any changes in protein expression of the PLCγ enzymes with decreasing temperatures, except for PLCγ1Δ20-22” and PLCγ2Δ20-22, which showed increased expression. It seems possible on the basis of these findings that the regions of PLCγ1 corresponding to the regions deleted in PLCγ2 in PLAID exert a stronger autoinhibitory influence on PLCγ1 than their PLCγ2 counterparts on PLCγ2 and that this difference blunts the stimulatory response of PLCγ1Δ19” and PLCγ1Δ20-22” to cooling. This possibility notwithstanding, the results indicate that the temperature sensitivities of PLCγ1 and PLCγ2 carrying deletions corresponding to exons 19 and 20-22 of PLCG2 are distinct.

The region encoded by exons 20 to 22 of PLCG2 contains an octapeptide (YRKMRRLRY) at the end of SH2c that is absolutely conserved in PLCγ1 and has previously been shown to mediate inhibition of PLCγ2 in trans and in cis [30,31]. To determine whether deletion of the octamer, designated PCI (for phospholipase C inhibitor), is sufficient to mediate sensitivity of PLCγ2 to cool temperatures, the deletion mutant PLCγ2ΔPCI was functionally compared to PLCγ2Δ20-22 and wild-type PLCγ2 (Fig. 4B, left panel). In addition, three other mutants carrying deletions known to promote constitutive activity of PLCγ2 at 37 °C, PLCγ2ASA, PLCγ2ASH, and PLCγ2ASH2c, were characterized (Fig. 4C, right panel). PLCγ2ΔPCI was analyzed again in the latter experiment as a control to ensure that the results shown in the two panels of Fig. 4B are comparable. Figs. 4B and S1B show that PLCγ2ΔPCI shared most features of its temperature sensitivity with PLCγ2Δ20-22 and, by extension, with PLCγ2Δ19 (cf. Fig. 1B): only a limited enhancement of its activity at 37 °C, a marked activation upon lowering the incubation temperature from 37 °C to 31 °C, and a decrease in activity at...
temperatures below 31 °C. In contrast, PLCγ2ΔSA, PLCγ2ΔSH, and PLCγ2ΔSH2c, although exhibiting constitutive activity at 37 °C, showed an only modest, further increase in activity with cooling, which was monophasic for PLCγ2ΔSH2c and only vaguely biphasic for PLCγ2ΔSA and PLCγ2ΔSH.

At first sight, the constitutive activities of the mutants PLCγ2ΔSA, PLCγ2ΔSH, and PLCγ2ΔSH2c observed at 37 °C in Fig. 4B, right panel, appeared rather limited in comparison to the constitutive activities reported earlier for similar mutants of PLCγ1 and PLCγ2 [31,32]. Therefore, the activities at 37 °C of wild-type PLCγ2, PLCγ2Δ19, PLCγ2Δ20-22, PLCγ2ΔPCI, PLCγ2ΔSH2c, PLCγ2ΔSH, and PLCγ2ΔSA were compared as a function of their expression levels in a more comprehensive analysis (Figs. 5A and B). The results show that, taking the relative abundance of the PLCγ2 variants into account (Fig. 5B), PLCγ2ΔPCI, PLCγ2ΔSA, PLCγ2ΔSH2c, and PLCγ2ΔSH exhibited higher constitutive activity at 37 °C than PLCγ2Δ19 and PLCγ2Δ20-22.

The fact that either of the two PLCγ2 mutants, PLCγ2Δ19 and PLCγ2Δ20-22, were activated by cool temperatures to a similar degree raised the question as to the effect of a combined deletion of the residues encoded by exons 19 through 22. Fig. 6 shows that, at 37 °C, the compound deletion mutant PLCγ2Δ19-22 displayed a markedly enhanced activity in comparison to either PLCγ2Δ19 or PLCγ2Δ20-22 alone, showing only marginal enhancements of their basal activities. The stimulatory effect of cooling to 31 °C was assayed at two expression levels of the deletion mutants. At both expression levels, the activity of PLCγ2Δ19-22 at 31 °C was much higher than the sum of the activities of PLCγ2Δ19 and PLCγ2Δ20-22. Hence, at both temperatures, 37 °C and 31 °C, and at the expression levels tested in this experiment, the deletions of PLCG2 exons 19 and 20-22 synergized to promote activation of PLCγ2.

3.4. The mouse PLCγ2 mutant Ali5 is also sensitive to activation by cool temperatures.

The symptoms observed in the mouse Ali5 and Ali14 disease models bear resemblance to the PLAID phenotype in some, but not in all respects [15,16,19,20]. In transfected COS-7 cells, the corresponding PLCγ2 mutants, PLCγ2Ali5 and PLCγ2Ali14 displayed only slightly enhanced basal activity at 37 °C, whereas the compound mutant, PLCγ2Ali5/Ali14 clearly showed constitutively enhanced activity [17]. In Ali5 mice, autoimmune inflammatory dermatitis commenced in the superficial layers of the paws and ears, i.e. in cool body regions [15]. This distribution resembles those of cutaneous granulomatous lesions in PLAID patients [19]. We therefore compared the effect of cool temperatures on the activity of the mutants PLCγ2Ali5, PLCγ2Ali14, and the compound mutant PLCγ2Ali5/Ali14. Figs. 7, left panel, and S2A show that all three mutants exhibited moderately enhanced activity at 37 °C (PLCγ2Ali5/Ali14 >
PLCγ2^{Ali5} > PLCγ2^{Ali14}) as described before [17]. Importantly, however, the three mutants displayed considerable differences in their sensitivities to cool temperatures, both in quantitative and in qualitative terms. Thus, while the response of PLCγ2^{Ali5/Ali14} to decreasing temperatures closely resembled the pattern observed for PLCγ2Δ20-22 (Figs. 7, center panel, and S2B), PLCγ2^{Ali14} showed only a minor, monophasic increase in activity, which was opposite to the decrease observed for wild-type PLCγ2 (Figs. 7, right panel, and S2C). PLCγ2^{Ali5} displayed an intermediate phenotype, nonetheless showing an almost 7-fold activation upon cooling from 37 °C to 31 °C (Fig. 7, right panel). These results are surprising since the Plcg2 mutations underlying the Ali5 and Ali14 phenotypes cause point mutations at positions outside of the SH2n-SH2c-SH3 region. Perhaps, similar structural and, hence, functional alterations are brought about by distinct mutations of the enzyme. Nevertheless, the results suggest that some of the lesions noticed in the affected animals, such as inflammation in superficial skin layers, may in fact be caused by further activation of PLCγ2^{Ali5} by cool temperatures. An intriguing question to be clarified by future investigation is whether PLCγ2 activated by other means, e.g. by other mutations or by tyrosine phosphorylation, are also sensitive to cold temperatures.

A more comprehensive comparison of the effects of the various constituents of the SH2n-SH2c-SH3-spPHc region, alone or in combination, on PLCγ2 constitutive activity and sensitivity to cool temperatures is shown in Fig. 8A. Unlike observed previously [31,32], deletion of SH3 also caused an increase in basal activity, which was even slightly higher (approximately 4.1-fold) than that monitored for deletion of SH2c (2.6-fold), but lower than that for deletion of all three SH domains (approximately 7.5-fold). The increases for the variants lacking two SH domains, PLCγ2ΔSH2nSH2c, PLCγ2ΔSH2cSH3, and PLCγ2ΔSH2nSH3, were approximately 3.6-, 1.9-, and 2.4-fold, respectively. While deletion of SH2n did not change the temperature sensitivity of PLCγ2 relative to the wild-type enzyme and deletion of all three SH domains resulted in a PLCγ2 mutant largely insensitive to cool temperature, all other deletion mutants were clearly sensitive. Maximal sensitivity (approximately 6.1-fold) was observed for PLCγ2ΔSH2cSH3, followed, in that order, by ΔSH2nSH3, ΔSH2nSH2c, ΔSH2c, and ΔSH3. Note that the degrees of stimulation by cool temperatures was by far lower for these mutants than the degrees observed for the PLAIΔ PLCγ2 mutants Δ19 and Δ20-22 (cf. Fig. 1). Fig. 8B shows a schematic representation of the deletions within the SH2n-SH2c-SH3 region (aa 515-840).
3.5. Cool-temperature-triggered activation of PLCγ2 deletion mutants is controlled by, but does not necessarily require the split PH domain.

PLCγ2 is activated by tyrosine phosphorylation and by activated Rac GTPases. The experiment shown in Fig. 9A was designed to determine the effect of decreasing temperatures on the ability of constitutively active Rac2G12V to activate PLCγ2Δ19 in comparison to wild-type PLCγ2. At 37 °C, Rac2G12V caused similar enhancements of the activity of both enzymes (Figs. 9A, left panel, and S3A). Lowering the incubation temperature, however, took a very different effect on Rac2G12V-mediated activation of the two enzymes. While only minor changes were observed for wild-type PLCγ2, there was a progressive loss of the stimulatory effect of Rac2G12V on PLCγ2Δ19. This loss is unlikely to be due to exhaustion of the inositol phospholipids substrate at lower temperatures, since PLCγ2Δ19 is well capable of producing even higher levels of inositol phosphates when expressed at higher density (Fig. 9A, right panel, and S3B). Likewise, it was unlikely that Rac2G12V became limiting at low temperatures, since stimulation of wild-type PLCγ2 was well retained even at the lowest temperature, 27 °C.

The activation of PLCγ2 by Rac is mediated by the internal, split PH domain of the enzyme [10,24]. The interdependence of the stimulatory effects of Rac2G12V and cooling evident in Figs. 9A and S3 prompted us to determine the role of this domain in PLCγ2 deletion mutant activation by cooling. Alanine replacement in spPH of PLCγ2 of W899, which is conserved in all PH domains [33] has previously been shown to result in a loss of PLCγ2 stimulation by Rac, but not by tyrosine phosphorylation [31]. Figs. 9B and S4A show that the W899A replacement completely abrogated the response of the enzyme to cool temperature activation. Furthermore, replacement of one or the other half of the split PH domain of PLCγ2 by the corresponding portions of PLCγ1, previously shown to block activation of the mutant enzymes by Rac, but to take no effect on their catalytic activity [24], also eliminated enzyme activation by cool temperatures. In contrast, replacement of both PH domains halves, which does not convey Rac sensitivity to PLCγ2 [24], fully rescued the response of PLCγ2 to cooling. These findings indicate that there is important structural interaction between the SH2n-SH2c-SH3 region and the surrounding sPH sequence. Two residues of the N-terminal half of the PLCγ1 spPH domain, Y509 and F510 (cf. Fig. 9C), have been shown to be important for the ability of spPHn to cooperate with SH2c to maintain PLCγ1 in an inactive conformation [17,34]. To examine the role of this site in the cool temperature response of PLCγ2, the two residues were substituted by alanine residues in PLCγ2Δ19 carrying the entire spPH of PLCγ1 (PLCγ2Δ19-PH11), thus generating the mutant PLCγ2Δ19-PH11-Y509A/F510A. Consistent with earlier findings [17,34], there was a slight increase in basal activity of the latter mutant in comparison to PLCγ2Δ19-PH11. Remarkably,
however, and in striking contrast to PLCγ2Δ19-PH11, there was no response of the mutant to cooling. Y509 and F510 of PLCγ1 corresponds to Y495 and C496 of PLCγ2. Most interestingly, alanine replacement of either one of the latter residues within the PLCγ2Δ19 (carrying its own, authentic spPH domain), caused a distinct loss of the enzyme’s response to cooling, replacement of both Y495 and C496 led to an almost complete loss of this response (Figs. 9D and S4B). These results indicate that a functional split PH domain, not necessarily the one genuinely contained in PLCγ2, is required with the context of the PLAID PLCγ2 deletion mutants to mediate their activation by cool temperatures.

The observation that PLCγ2ΔASH, a mutant lacking the entire SH2n-SH2c-SH3 region, displays enhanced basal activity, but is largely insensitive to activation by cold is remarkable, because this mutant lacks all constituents thought to mediate autoinhibition of the enzyme (Fig. 8B). On the one hand, this finding clearly suggests that the activation pattern of the two PLAID mutants, PLCγ2Δ19 and PLCγ2Δ20-22, is distinct from a mere loss of autoinhibition, and indicates, by extension, that it is based on a specific molecular mechanism(s). On the other hand, the observation is rather puzzling, because cold activation appears to be, at the same time, dependent on and independent of specific constituents of the SH2n-SH2c-SH3 region. However, given the importance of the split PH domain for cold activation of PLCγ2 emerging from the experiments shown in Fig. 9B and D, the possibility remained that activation by cold temperatures is sterically and/or conformationally restricted in the mutant PLCγ2ΔASH by the (artificial) covalent linkage between the two halves of the split PH domain (cf. Fig. 9C). To examine this possibility, we took advantage of earlier findings suggesting that PLC isozymes can be expressed as two independent polypeptide fragments containing either of the two catalytic subdomains, X and Y [35,36]. Thus, PLCγ2ΔASH was expressed either as a single polypeptide and as two separate chains, encompassing either of the two catalytic subdomains XPHc and YPHc, either one containing the corresponding half of the split PH domain, followed by functional analysis at 37 °C and 31 °C. Figs. 10A and S5 show that expression of the N-terminal or the C-terminal half of PLCγ2 did not lead to enhanced inositol phosphate formation. In marked contrast, when both halves, XPHn and YPHc, were coexpressed, basal inositol phosphate formation at 37 °C was markedly enhanced. More importantly, however, decreasing the incubation temperature to 31 °C caused a substantial enhancement of PLC activity, in striking contrast to cells expressing the ΔASH deletion mutant of PLCγ2, where no change in activity was observed. These results strongly suggest that the structural element(s) mediating cold activation of PLAID PLCγ2 mutants reside outside the SH2n-SH2c-SH3 region and that its responsiveness to cool temperatures is blocked by covalent linkage between the two portions of the PLCγ2 split PH domain.

To examine the requirement of the split PH domain in itself for cool-temperature-mediated activation of coexpressed XPHn and YPHc, the coding regions of the split PH domain halves were
removed from the two fragments, resulting in fragments X and Y. The two fragments were expressed alone or together and functionally analyzed at 31 °C or 37 °C. Most intriguingly, in contrast to cells expressing either X or Y alone, cells coexpressing fragments X and Y displayed a marked enhancement of inositol phosphate formation by lowering the incubation temperature to 31 °C (Fig. 10B). This enhancement was similar in extent as the one observed for PLCγ2Δ20-22 (approximately 32-fold versus 41-fold). Thus, although the split PH domain appears to control the cool temperature sensitivity of the PLAID PLCγ2 mutants, it is not required per se for cool-temperature-mediated activation of the enzyme’s core constituents. The expression of wild-type and Δ20-22 mutant PLCγ2 as well as fragments X and Y was similar at 31 °C and 37 °C, respectively (Fig. S6).

3.5. The PLAID PLCγ2 mutants are resistant to activation by EGF receptors endogenously expressed in COS-7 cells.

The rather puzzling concurrence of gain and apparent loss of function of PLCγ2 in certain PLAID patient cells [19,37] prompted us to determine the effects of the two PLAID PLCγ2 mutations on the response of the mutant enzymes to tyrosine kinase receptor activation. To this end, we made use of EGF receptor tyrosine kinases endogenously expressed in COS-7 cells and capable of both regulating several endogenous signalling functions and activating exogenous PLCγ2 [38,39,17] and examined their ability to stimulate PLCγ2-mediated inositol phosphate formation. Fig. 11 shows that addition of 100 ng/ml of EGF caused similar (approximately 12-fold) increases in wild-type PLCγ2 activity at 37 °C and 31 °C. In contrast, neither PLCγ2Δ20-22 nor PLCγ2Δ19 were affected by addition of EGF, regardless of whether the radiolabeling of cells and their treatment with EGF was done at 37 °C or 31 °C. The inability of EGF to mediate PLCγ2 PLAID mutant stimulation at 37 °C was not due to lack of available enzyme substrate, since the two mutants were markedly sensitive to activation by cooling to 31 °C. Additional experiments (not shown) revealed that substrate depletion did also not occur in the presence of the mutant PLCγ2 enzymes at 31 °C. Fig. S7 shows that the functional changes depicted in Fig. 11 were not due to changes of wild-type or mutant PLCγ2 expression.

4. Discussion

The results presented in this work show that deletion of residues encoded by exons 19 and 20-22 from PLCγ2 causes both a mild constitutive activation of inositol phosphate formation at 37 °C and a marked, several-hundred-fold enhancement of this activity in response to very small decreases in temperature upon expression of the mutant enzymes in intact cells. These functional responses are qualitatively and quantitatively similar, if not identical for both types of mutants. While the former changes are consistent with those reported earlier for transfected cells, the latter are much higher than
the small increases in [Ca$^{2+}$], observed when peripheral blood B cells of PLAID patients are exposed to cold. In the latter case, differences to normal B cells were only observed at temperatures below 29 °C, not exceeding a maximum of an about two-fold increase at 21 °C [19]. The results shown here suggest that the primary defect of the mutant enzymes, i.e. sensitivity to stimulation by cool temperatures, emerges at temperatures only a few degrees below the normal body temperature. These findings are in agreement with clinical findings on patients with a deletion of exons 20-22, where only very subtle cooling, such as the one caused by a single tear rolling down the cheek of an affected family member at room temperature caused symptoms within one minute [20]. In additional experiments (not shown), we found that application of single drops of water (60 µl) to human skin caused a transient, evaporative-heat-loss-mediated decrease in skin temperature by about 6 °C. Of note, even at an ambient, comfortable temperature of 27 °C, the temperature in many cutaneous and subcutaneous regions is already somewhat lower than the esophageal core temperature of 37 °C [40], such that only minimal further temperature decreases by evaporative heat loss may suffice to cause maximal PLC$\gamma_2$ activation. The considerable decrease of the stimulatory effect observed at temperatures below 31 °C (Fig. 3) may explain, together with the decrease in inositol phospholipid synthesis (Fig. 1D) - why PLAID patients are typically negative in the cold stimulation time test (CSTT), involving cooling with ice, and upon cold-water immersion [19,20]. Under these conditions, the cooled tissues may reside at the enzyme-activating temperatures only very transiently, i.e. too shortly for allowing effective activation mutant PLC$\gamma_2$. The time course of PLC$\gamma_2$ deletion mutant activation and its reversible nature closely matches the clinical observation on PLAID patients of an immediate onset of inflammatory symptoms upon evaporative skin cooling and a somewhat slower resolution of the symptoms developing over the course of 30 minutes of rewarming at room temperature [20]. Mouse PLC$\gamma_2$ carrying a gain-of-function mutation, D993G, in the catalytic region, PLC$\gamma_2^{Ali5}$, and causing spontaneous autoinflammation and autoimmunity shows a similar, albeit less dramatic response to cold temperatures. This suggests that some of the lesions noticed in the affected animals, such as inflammation in superficial skin layers, may in fact be caused by further activation of PLC$\gamma_2^{Ali5}$ by cool temperatures. An intriguing question to be clarified by future investigation is whether PLC$\gamma_2$ activated by other means, e.g. by other mutations [21] or by tyrosine phosphorylation, are also sensitive to cold temperatures.

Three mechanisms, not necessarily mutually exclusive, should be considered to explain the sensitivity of the PLC$\gamma_2$ mutants to cold temperatures in intact cells. First, other cold-sensitive molecules endogenously present in COS-7 cells - and, possibly, in mast cells - might indirectly cause activation of the mutants, e.g. via a soluble mediator or via altered protein-protein-interaction patterns. Second, the mutants may be specifically enabled to sense temperature-mediated changes in the physical properties of the plasma membrane phospholipid bilayer containing the enzyme’s
substrate(s). The third possibility is that an intrinsic conformational change is induced in mutant PLC\(\gamma_2\) by temperatures between 37 °C and 31 °C. With regard to the first possibility, we would like to point out that, among the known cold-temperature-sensitive molecules, TRPA1, TRPM8, and TRPC5, the former two responded to cold temperatures only within lower temperature ranges [23] and endogenous expression of the latter two was not evident in COS-7 cells [39-44]. While we cannot at present formally exclude the second mechanism, we note that the phase transition temperature (\(T_m\)) of dipalmitoylphosphatidyl-choline (the most representative lipid in model membrane studies) in artificial phospholipid membrane vesicles is 41 °C [45] and thus outside the range of temperatures mediating activation of the PLC\(\gamma_2\) mutants. However, the behavior of native plasma membranes may be different. More experimentation is required to elucidate the exact nature of the interplay among the mechanisms potentially involved in cool-temperature-mediated activation of PLAID PLC\(\gamma_2\) mutants under conditions of physiological PtdIns(4,5)\(P_2\) presentation as a substrate to PLC\(\gamma_2\) in native plasma membranes of intact cells.

If the structural element mediating, directly or indirectly, the sensitivity of PLC\(\gamma_2\) deletion mutants to cool temperatures in intact cells resides on the enzyme itself, the simplest interpretation of the results would be that removal of the portions encoded by exons 19 or 20-22 causes a loss of an autoinhibition, since the two regions overlap with regions identified before [11,12,31,32] or in this study (cf. Fig. 8) to be autoinhibitory at 37 °C. However, the results shown in Fig. 8 suggest that constitutive activity at 37 °C and stimulation by cool temperatures are not necessarily correlated with each other. While this would not formally exclude a loss of autoinhibition, acquisition of an autostimulatory mechanism sensitive to decreasing temperatures appears as an alternative explanation. At first glance, a structural element residing in the SH domain region of PLC\(\gamma_2\) would appear a likely candidate for triggering such a stimulation. However, the deletion experiments shown in Fig. 8A revealed that none of the constituents of the entire SH2n-SH2c-SH3 region is specifically required for enzyme activation by cool temperatures and that even a deletion mutant lacking both of the short partial repeats shared between the two SH2 domains (PLC\(\gamma_2\)\(\Delta\)SH2nSH2c) is activated by incubation at 31 °C.

Thus, the possibility remains that cool-temperature-sensitivity in intact cells resides either in the split PH domain or in the remainder of the enzyme, and that this sensitivity is specifically kept in check by either the intact, native SH region, by its SH2c-SH3 portion, or by a covalent linkage between the two split PH domain halves. At first sight, the fact that the cool-temperature-sensitivity is lost when non-homonymous split PH domain halves are present in PLC\(\gamma_2\), when the tertiary structure of the PH domain is disrupted by the W\(^{899}\)A mutation [17], or when two residues of its N-terminal half, Y\(^{509}\) and F\(^{510}\), are replaced by alanine residues, supports an important regulatory role of the split
PH domain. The latter residues have recently been mapped, by NMR titration analysis, to the interface of spPH and SH2c and suggested to be involved in SH2c-mediated autoinhibition [11]. The fact that removal of the two residues is also effective in the absence of almost the entire SH2c domain in PLCγ2Δ19 is difficult to reconcile with an important role of a spPH-SH2c interaction in cool-temperature regulation of mutant PLCγ2. However, chemical shift perturbations observed in NMR experiments probing protein-protein-interactions may imply structural reorientation of residues upon binding rather than direct involvement in the interaction surface [46]. This and the weak interaction between the two domains observed before [32] is consistent with additional functions of these two spPH domain residues. Interestingly, Y509 of PLCγ1 corresponds to Y495 of PLCγ2, which is mutated to C495 in PLCγ2Ali14 and the double mutant PLCγ2Ali5/Ali14. The considerable stimulatory effect of the Ali14 mutation on the stimulation of PLCγ2Ali5 by cool temperature (cf. Fig. 7, left panel) suggests there may loss- and gain-of-function mutations in this locus with regard to cool-temperature sensitivity of PLCγ2. Collectively, the results discussed up until this point suggest a new regulatory role of the PLCγ2 split PH domain in mutant enzyme activation by cool temperatures. However, and quite surprisingly, the results shown in Fig. 10B strongly suggest that the split PH domain is not required as such for cool-temperature-mediated activation of mutant PLCγ2.

A model summarizing and schematically conceptualizing the responses of the PLAID PLCγ2 and their further truncated variants to cooling from 37 °C to 31 °C is shown in Fig. 10C-H. We would like to point out that these schematic views are limited by the fact that they solely rely on measurements of PLCγ2 enzyme activity and that they await confirmation by direct structural analysis or by other types of independent experimental evidence. Nevertheless, at least for the PLAID PLCγ2 mutants, our results suggest a dynamic, regulatory role of spPH in controlling the cool temperature response of these mutants. It is tempting to speculate that the interaction of spPH with activated Rac is lost during that response (cf. Fig. 9A). Of interest, the spPH domains of PLCγ have been suggested before to undergo dynamic changes leading to formation of intermolecular PH domains regulating agonist-mediated Ca2+ entry into cells [47-49], although this has remained somewhat controversial [9].

The marked sensitivity of the two PLAID PLCγ2 mutants to cool temperatures may explain the gain-of-function symptoms observed in PLAID patients such as cold urticaria [19,20] and skin granulomas [50]. They are likely to be caused by cool-temperature-mediated activation of PLCγ2 in cutaneous mast cells, neutrophils, and monocytes, where the enzyme plays important roles in FceR-, integrin-, and FcγR-mediated inflammatory skin reactions [51,52]. Symptomatic allergic and autoimmune disease, occurring in 56 % and 26 % of PLAID patients and potentially precipitated by enhanced basal (37 °C) or cool-temperature-mediated activation of PLCγ2 in mast and other immune cells, may also fall into this category. The latter may include B cells and dendritic cells. Loss-of-
function symptoms, such as antibody deficiency, recurrent sinopulmonary infection, and symptoms resembling certain forms of common variable immunodeficiency [53], detected in 75%, 44%, and 11%
respectively, of PLAID patients, are more difficult to explain. However, at least in the heterologous
expression system used here, the two PLAID mutants appeared to be resistant to stimulation by
activation of endogenous EGF receptors, both at cool (31 °C) and physiologic (37 °C) temperatures
(Fig. 11). These results suggest that resistance to receptor stimulation may, at least in part, be an
intrinsic property of the PLAID mutant enzymes themselves or of components endogenously present
in COS-7 cells. This is an important issue, since we expect, but certainly need to prove, that
observations reported herein in transfected COS-7 cells will be relevant for, e.g., B cells with
endogenous PLCγ2 expression. As for the B cell context, recent results suggest that the SH2c domain
of PLCγ2 plays a critical role in stabilizing the early BCR signaling complex, such that PLCγ2 mutants
lacking a functional cSH2 domain such as PLCγ2Δ19 and PLCγ2Δ20-22 may act in a dominant-
negative manner to prevent the formation of stable, signaling-competent BCR clusters consisting of
Syk, BLNK, Btk, and PLCγ2 [37]. Furthermore, association of BCR with the inhibitor of Ca2+
signaling, Cbl, was dysregulated in B cells expressing PLCγ2Δ20-22. These results notwithstanding, it
is well known that constant antigen receptor occupancy and signaling, including an increased basal
concentration of intracellular Ca2+ ([Ca2+]i basal) mediates maintenance of B cell anergy, which is
characterized by the persistence in the periphery of B cells unresponsive to immunogen [54-58]. A
defect leading to impaired BCR-induced increases in [Ca2+]i has been described for type Ia patients
with common variable immunodeficiency (Freiburg classification) and shown to be associated with a
reduction in IgD IgM+ CD27+ class-switched memory B cells, hypogammaglobulinemia, and
autoimmune dysregulation [59], all of which are observed in PLAID patients. In normal B cells,
anergy is rapidly reversed after dissociation of self antigen, e.g. by using hapten competition, and
these cells regain antigen responsiveness [52]. In PLAID B cells, [Ca2+]i basal would be expected to be
elevated, either chronically due to constitutively enhanced PLCγ2 mutant activity at 37 °C or
intermittently during passage through cool body regions of the patient, causing reversible reductions of
B cell responsiveness. Some of the trans-inhibitory mechanisms observed in anergic B cells, inhibiting
the signaling of G-protein-coupled chemoattractant receptors [59], may be operative even in other
immune cells, such as neutrophils, potentially contributing to the loss-of-immune-response
phenotype, e.g. reduced directed migration, observed in PLAID patients [50]. Anergic mechanisms
are also in effect in human mast cells in response to persistent cell activation, changing the cells’ set
points for further activation [60]. These mechanisms could explain why cold urticaria is not generally
observed in PLAID patients in all regions where the temperature on the skin should be low enough to
cause mutant PLCγ2 activation [37]. In human skin, most mast cells are present immediately below the
dermo-epidermal junction [61] and are, hence, exposed to temperatures very similar to those
prevailing on the skin. Small negative deviations from the set point temperature developing over short
time periods may be the relevant trigger of cold urticarial lesions in PLAID patients.

An interesting question raised by the dramatic functional consequence of the *PLCG2* deletions
observed in PLAID patients is whether the genomic regions encompassing exons 19 through 22 are
subject to alternative processing of pre-messenger RNA, leading to exclusion of residues encoded by
exons 19 and/or 20-22 from the mature PLCγ2 protein. This appears as an important issue, since
almost 90% of human genes undergo alternative splicing with a minor isoform frequency of 15% or
more, with variations, intraindividually, between tissues and between individuals [62]. Alternative
RNA splicing has been shown to be pervasive across immune system lineages [63]. Intriguingly, one
of the two changes observed for PLCγ2-encoding mRNA in mouse CD19+ B cells is skipping of exons
20-22. The marked similarity between the genomic organization of the mouse *Plcg2* and the human
*PLCG2* genes raises the possibility that this alteration may also exist in humans to convey cool-
temperature-sensitivity to PLCγ2 in certain cell types and/or particular individuals. Cold environmental
temperatures are associated with a number of human disease states, including, e.g., exercise-induced
asthma and acute cold-induced coronary events [64,65]. Cold-mediated activation of variant PLCγ2 in
these states, e.g. in mast cells, appears as an intriguing possibility.

4. Conclusions

(i) The two human PLAID PLCγ2 mutants PLCγ2Δ19 and PLCγ2Δ20-22 are strongly (> 100-fold),
rapidly, and reversibly activated in intact cells by cooling the cells by only a few degrees. (ii) The
underlying mechanism(s) is distinct from a mere loss of SH-region-mediated autoinhibition and
dependent on both the integrity and the pliability of the spPH domain. (iii) The results suggest a new
mechanism of PLCγ activation with unique thermodynamic features and assign a novel regulatory role
to its spPH domain.

Supplementary data to this article can be found online.
Funding

Work in P.G.’s laboratory is funded by grants from the Deutsche Forschungsgemeinschaft (DFG) (SFB 1074, TP A8). The contribution of J.K. and H.K. was supported by project Z1 of SFB 1074. A.S. was a fellow of the DFG Research Training Group GRK1041 and the International Graduate School in Molecular Medicine Ulm (IGradU) funded within the Excellence Initiative of the German Federal and State Governments.

Author contributions

A.S., C.W., M.W., J.H., P.V., J.K., D.F., H.K., and G.H. performed the experiments and analyzed the data. P.G. provided overall direction and wrote the manuscript with input from J.M. and the other authors.

Conflict of interest

The authors declare that they have no conflicts of interest with the contents of this article.

Acknowledgements

The expert technical assistance of Susanne Gierschik and Norbert Zanker is greatly appreciated. Special gratitude goes to Armin Buehler for custom designing and constructing the incubation chambers. We are grateful to Drs. Bertil Hille, Tibor Rohacs, Eleonora Zakharian, Hal M. Hoffman, and Nicholas G. Kounis, and to Elisabeth Hermkes for very helpful discussions.
References


**Figure legends**

**Fig. 1.** The PLC\(\gamma_2\) deletion mutants PLC\(\gamma_2\)Δ19 and PLC\(\gamma_2\)Δ20-22 are specifically activated by cool temperatures. (A) Domain organization of the PLC\(\gamma\) isoforms. The positions of the two deletions Δ19 and Δ20-22 are indicated. *aa*, amino acids. (B) COS-7 cells were transfected with 500 ng each per well of either empty vector (■), or vector encoding either wild-type PLC\(\gamma_2\) (●), PLC\(\gamma_2\)Δ19 (▲), or PLC\(\gamma_2\)Δ20-22 (▲). Twenty-four hours after transfection, the cells were incubated for 20 h with myo-[2-\(^3\)H]inositol at the indicated temperatures and inositol phosphate formation was then determined. The levels of inositol phosphate formation at 37 °C are shown in expanded scale on the right vertical axis (open symbols). (C) Expression of wild-type and mutant PLC\(\gamma_2\) isozymes in the experiment shown in Fig. 1B. Cells from one well each were washed once with 0.2 ml of Dulbecco’s PBS and then lysed by addition of 100 µl of SDS-PAGE sample preparation buffer. The samples were subjected to SDS-PAGE and immunoblotting was performed using an antibody reactive against the c-Myc epitope on PLC\(\gamma_2\). (D) The activation of PLC\(\gamma_2\) deletion mutants by cool temperatures is not explained by changes in cellular levels of inositol phospholipids. COS-7 cells were transfected with 2 µg each per well of empty vector (*Control*), vector encoding wild-type PLC\(\gamma_2\) (*WT*), or PLC\(\gamma_2\)Δ19 (*Δ19*). Twenty-four hours after transfection, the cells were incubated for a further 20 h in individual incubation chambers with myo-[2-\(^3\)H]inositol at the indicated temperatures. The amount of [\(^3\)H]inositol present in the culture medium after radiolabeling of the cells (■) and the cellular formation of inositol phospholipids (●) was then determined as described in Experimental Procedures. The data was analyzed by non-linear least square curve fitting to a polynomial second order (quadratic) equation. (E) The expression of wild-type PLC\(\gamma_2\) and PLC\(\gamma_2\)Δ19 was examined by subjecting cells from one well each to analysis by SDS-PAGE and immunoblotting using antibodies reactive against the c-Myc epitope.

**Fig. 2.** The activation of PLC\(\gamma_2\)Δ19 and PLC\(\gamma_2\)Δ20-22 by subphysiological temperatures occurs after protein synthesis and is reversible. (A) COS-7 cells were transfected with 500 ng each per well of either vector encoding wild-type PLC\(\gamma_2\), PLC\(\gamma_2\)Δ19, or PLC\(\gamma_2\)Δ20-22. Twenty-four hours after transfection, the cells were incubated for a further 20 h at 31 °C or 37 °C in individual incubation chambers with myo-[2-\(^3\)H]inositol, as indicated in the absence or presence of 100 µg/ml cycloheximide (*CHX*). (B) Expression of wild-type and mutant PLC\(\gamma_2\) isozymes in the experiment shown in panel A. The control samples (*Co.*) were taken at the end of the initial 24-h-transfection phase. Cells from one well each were analyzed by SDS-PAGE and immunoblotting using an antibody
reactive against the c-Myc epitope. We note that despite its effect on PLC$_{\gamma2}$ protein abundance, cycloheximide had only a minor effect on the increased formation of inositol phosphates by the two deletion enzymes at 31 °C versus 37 °C (panel A). We think that this is due to the fact that the upper limits of available phospholipid substrate were reached in this experiment. Hence, the levels inositol phosphate formation in the samples containing the deletion mutants assayed at 31 °C may have exceeded those determined in panel A under conditions of unlimited substrate supply. (C) COS-7 cells were transfected with 500 ng each per well of either vector encoding wild-type PLC$_{\gamma2}$, PLC$_{\gamma2}\Delta19$, or PLC$_{\gamma2}\Delta20-22$. Twenty hours after transfection the cells were pre-incubated for four hours in individual incubation chambers at 31 °C (31→31; 31→37) or 37 °C (37→37). The cells were then incubated for another 20 h with myo-[2-3$^3$H]inositol in individual incubation chambers at 31 °C (31→31) or 37 °C (31→37; 37→37). (D) Expression of wild-type and mutant PLC$_{\gamma2}$ isozymes in the experiment shown in panel C. (E) Time course. COS-7 cells were transfected with 500 ng each per well of either vector encoding wild-type PLC$_{\gamma2}$ (●), PLC$_{\gamma2}\Delta19$ (▲), or PLC$_{\gamma2}\Delta20-22$ (▼). Cells were transfected and radiolabeled and inositol phosphate formation was then determined. At the indicated time points, the cells were shifted from 37 °C to 31 °C and back to 37 °C, respectively. Open symbols, incubation at 37 °C throughout. (F) Expression of wild-type and mutant PLC$_{\gamma2}$ isozymes in the experiment shown in panel E. Cells from one well each were analyzed by SDS-PAGE and immunoblotting using an antibody reactive against the c-Myc epitope.

**Fig. 3.** Determination of the 10-degree temperature coefficients, $Q_{10}$. The data shown in Fig. 1B on the cool temperature responses of cells expressing wild-type PLC$_{\gamma2}$ (WT), PLC$_{\gamma2}\Delta19$ (∆19), and PLC$_{\gamma2}\Delta20-22$ (∆20-22) was taken to determine the $Q_{10}$ values of these responses as detailed in Experimental Procedures. The individual temperatures $T_i$ were plotted against $\log_{10}\left(\frac{A_i}{A_{ref}}\right)$, with the maximum activity of PLC$_{\gamma2}\Delta19$ at 31 °C chosen as the reference activity $A_{ref}$ and reference temperature, $T_{ref}$, respectively. The data of the linear components was analyzed by non-linear least square curve fitting to a polynominal first order (straight line) equation. The slopes of the curves of PLC$_{\gamma2}\Delta19$ and PLC$_{\gamma2}\Delta20-22$ were not significantly different by global curve fitting using shared parameters from each other and, between 25 °C and 31 °C, from the slope obtained for wild-type PLC$_{\gamma2}$ ($P = 0.9356$ and $P = 0.2121$, respectively).

**Fig. 4.** Cool-temperature-mediated activation of PLC$_{\gamma2}\Delta19$ and PLC$_{\gamma2}\Delta20-22$ is distinct from loss of SH-region-mediated autoinhibition. (A) The temperature sensitivities of the PLAI$_{D}$ PLC$_{\gamma2}$ deletion mutants are unique. COS-7 cells were transfected with 500 ng each per well of vector encoding either wild-type PLC$_{\gamma1}$ (●), PLC$_{\gamma2}\Delta19$ (▼), PLC$_{\gamma2}\Delta20-22$ (△) or mutants of PLC$_{\gamma1}$ carrying deletions corresponding to those in PLC$_{\gamma2}$ and referred to as PLC$_{\gamma1}\Delta19"$ (▼) and PLC$_{\gamma1}\Delta20-22"$ (△). (B)
The effects of deletions within the SH2n-SH2c-SH3 region on cool temperature regulation of PLCγ2 are specific. COS-7 cells were transfected with 50 ng each per well of vector encoding either wild-type PLCγ2 (●), PLCγ2Δ20-22 (▲), or PLCγ2ΔPCI (■) (left panel), or with 10 ng each per well of vector encoding either wild-type PLCγ2ΔSA (○), PLCγ2ΔSH (□), or PLCγ2ΔSH2C (open diamonds), or 50 ng each per well of vector encoding PLCγ2ΔPCI (■) as a control (right panel). The very similar responses of cells expressing PLCγ2ΔPCI shows that the results shown in the two panels are comparable.

**Fig. 5.** Basal activities of wild-type and mutant PLCγ2 isozymes at 37 °C. (A) COS-7 cells were transfected with either 500 ng each per well of empty vector (Co., control) or increasing amounts (10 ng, 150 ng, and 500 ng) of vector encoding wild-type PLCγ2 (WT), PLCγ2Δ19 (Δ19), PLCγ2Δ20-22 (Δ20-22), PLCγ2ΔPCI (ΔPCI), PLCγ2ΔSH2c (ΔSH2c), PLCγ2ΔSH (ΔSH), or PLCγ2ΔSA (ΔSA). Twenty-four hours after transfection, the cells were incubated for 20 h at 37 °C with myo-[2-3H]inositol and inositol phosphate formation was then determined. (B) Expression of wild-type and mutant PLCγ2 isozymes in the experiment shown in panel A. Cells from one well each were analyzed by SDS-PAGE and immunoblotting using antibodies reactive against the c-Myc epitope (upper panel) or β-actin (lower panel).

**Fig. 6.** The deletions of PLCG2 exons 19 and 20-22 synergize to promote basal and cool-temperature-mediated activation of PLCγ2. Left panel, COS-7 cells were transfected as indicated at the abscissa with 500 ng each per well of either vector encoding wild-type PLCγ2 (WT), or 500 ng, 10 ng, or 20 ng of vector encoding PLCγ2Δ19 (Δ19), PLCγ2Δ20-22 (Δ20-22), or PLCγ2Δ19-22 (Δ19-22). Twenty-four hours after transfection, the cells were incubated for a further 20 h, as indicated at the abscissa at 31 °C or 37 °C, in individual incubation chambers with myo-[2-3H]inositol. Inositol phosphate formation was then determined as described under Experimental Procedures. The significance of differences between means ± standard errors was assessed by using the unpaired t test with two-tailed P values with or without Welch’s correction as appropriate, as contained in GraphPad InStat®, version 3.10. Two effects are said to be synergistic, if the effect of two components tested in combination is statistically significantly higher than the sum (+) of the individual effects (**, 0.001 < P < 0.01; ***, P < 0.001). Right panel, COS-7 cells were transfected with 500 ng of vector encoding wild-type PLCγ2 (WT), PLCγ2Δ19 (Δ19), PLCγ2Δ20-22 (Δ20-22), or PLCγ2Δ19-22 (Δ19-22). Twenty-four hours after transfection, the cells were incubated for a further 20 h. Cells from one well each were then analyzed by immunoblotting using an antibody reactive against the c-Myc epitope.

**Fig. 7.** Effect of cool temperatures on the activities of PLCγ2Al5, PLCγ2Al14 and PLCγ2Al5/14. Left panel, COS-7 cells were transfected with 500 ng each per well of either empty control vector, or vector
encoding either wild-type PLCγ2, PLCγ2Alis, PLCγ2Alih, or PLCγ2Alis/Alih. Twenty-four hours after transfection, the cells were incubated for 20 h in individual incubation chambers with myo-[2-3H]inositol at 31°C or 37°C. Middle and right panel, COS-7 cells were transfected as indicated with 500 ng each per well of vector encoding either wild-type PLCγ2 (○), PLCγ2Δ20-22 (▲), PLCγ2Alis (■), PLCγ2Alih (●), or PLCγ2Alis/Alih (●). Twenty-four hours after transfection, the cells were incubated for 20 h in individual incubation chambers with myo-[2-3H]inositol at the indicated temperatures. In the right panel, cells that had been transfected with vector encoding PLCγ2Alis/Alih were also analyzed as a control. The response of these cells was very similar, both in qualitative and quantitative terms (not shown), to the response of the PLCγ2Alis/Alih-expressing cells shown in the left and in the center panel, indicating that the results shown in the three panels are comparable.

**Fig. 8.** Functional effects of deletions within the SH2n-SH2c-SH3 region. (A) COS-7 cells were transfected as indicated with 500 ng each per well of empty control vector or vector encoding wild-type PLCγ2, or with 10 ng each per well of vector encoding PLCγ2ΔSH2n, PLCγ2ΔSH2c, PLCγ2ΔSH3, PLCγ2ΔSH2nSH2c, PLCγ2ΔSH2cSH3, PLCγ2ΔSH2nSH3, or PLCγ2ΔSH. Twenty-four hours after transfection, the cells were incubated for a further 20 h in individual incubation chambers at 37 °C or 31 °C. (B) Schematic representation of the deletions within the SH2n-SH2c-SH3 region (aa 515-840). The positions of the two partial repeats detected within the two SH2 domains (KDGFLVR/RDGAFLIR and GRVQHCRIGKV/KHCRIG; SH2n/SH2c, identical residues underlined) and of the various deletions used in this study are indicated.

**Fig. 9.** Cool-temperature-triggered activation of PLAIδ PLCγ2 deletion mutants is mediated by the split PH domain. (A) Activation of PLCγ2Δ19 by Rac2G12V is diminished at cool temperatures. Left panel, COS-7 cells were cotransfected with 500 ng each per well of vector encoding either wild-type PLCγ2 (■,□) or 100 ng of vector encoding PLCγ2Δ19 (●,○) together with 25 ng per well of either empty vector (□,○) or vector encoding Rac2G12V (■,●). There was no effect on inositol phosphate formation upon expression of Rac2G12V in the absence of PLCγ2 isozymes (not shown). Right panel, COS-7 cells were transfected with either 500 ng per well of vector encoding wild-type PLCγ2 (○) or increasing amounts of vector encoding PLCγ2Δ19 (●). (B) The split PH domain mediates activation of PLCγ2Δ19 by cold. COS-7 cells were transfected with 500 ng each per well of vector encoding either wild-type PLCγ2, PLCγ2Δ19, PLCγ2Δ19W899A, PLCγ2Δ19-H12, PLCγ2Δ19-PH21, PLCγ2Δ19-PH11, PLCγ2-PH11, or PLCγ2Δ19-PH11Y509A/F510A. (C) Three-dimensional structures of the split PH domains of PLCγ1 [2FJL [9]] and PLCγ2 [2W2X [10]], as analyzed and visualized using the PyMOL Molecular Graphics System. The positions of the residues mutated in (B) and the site of insertion of the SH2n-SH2c-SH3 region in wild-type PLCγ2, covalently linked in PLCγ2ΔSH are indicated. (D) A functional
spPH domain is required for the temperature sensitivity of PLCγ2Δ19. COS-7 cells were transfected with 500 ng each per well of vector encoding either wild-type PLCγ2, PLCγ2Y495A, PLCγ2C496A, PLCγ2Y495A/C496A, PLCγ2Δ19, PLCγ2Δ19Y495A, PLCγ2Δ19C496A, or PLCγ2Δ19Y495A/C496A.

**Fig. 10.** Cool temperature sensitivity of PLCγ2ΔSH mutant is restored by expression of the enzyme as two separate chains. (A) COS-7 cells were transfected with 500 ng each per well of either empty vector, 500 ng of vector encoding wild-type PLCγ2, 10 ng of vector encoding PLCγ2ΔSH, or 250 ng of vector encoding PLCγ2-XPHn or PLCγ2-YPHc (both encompassing the respective halves of the split PH domain), or cotransfected with 250 ng of vector encoding PLCγ2-XPHn together with 250 ng of vector encoding PLCγ2-YPHc. (B) The split PH domain is not essential for the cool temperature sensitivity of PLCγ2 expressed as two separate chains. COS-7 cells were transfected with 500 ng each per well of either empty vector (Co.), or vector encoding wild-type PLCγ2 (WT), PLCγ2-X (X), or PLCγ2-Y (Y) (the latter two lacking the respective halves of the split PH domain) (all four vectors at 500 ng per well), or cotransfected with 100 ng each per well of vectors encoding PLCγ2-X or PLCγ2-Y. PLCγ2Δ20-22 (Δ20-22) (500 ng vector per well) was analysed for comparison. (C-H) Model of the activation of PLAID PLCγ2 mutants, exemplified by PLCγ2Δ20-22, and their deletion mutants by cool temperature. (C) Wild-type PLCγ2 is autoinhibited in its basal state by constituent(s) of its SH domain region and does not respond to cooling (-). (D) The Δ20-22 deletion within the SH2n-SH2c-SH3 region causes a reorientation of the SH domain region, allowing the enzyme to be markedly activated (++) by cool temperatures in a process depending on the integrity of spPH. Only minor functional changes are caused by the deletion at 37 °C (not shown). (E) Covalent linkage of the two PH domain halves in the constitutively active (+) deletion mutant PLCγ2ΔSA prevents further activation by cooling. (F) Disengaging the two spPH halves from the tight and stable covalent linkage allows cool temperature activation of the bipartite enzyme (++). (G and H) Coexpression of fragments X and Y yields enzyme that is constitutively active at 37 °C (at suitable expression levels, not shown) (+). Cooling to 31 oC results in marked further activation (++). Note that cool temperature activation of the bipartite PLCγ2 mutants occurs in the absence of any of the SH2n-SHc-SH3 constituents (F and H).

**Fig. 11.** The PLCγ2 deletion mutants Δ20-22 and Δ19 are resistant to stimulation by EGF. (A) COS-7 cells were transfected with vectors encoding either wild-type PLCγ2 (WT) (500 ng per well), PLCγ2Δ20-22 (Δ20-22), or PLCγ2Δ19 (Δ19) (both at 100 ng per well). Eighteen hours after transfection, the cells were incubated for a further 24 h as indicated at either 31 °C or 37 °C with myo-[2,3H]inositol and 10 mM LiCl in the absence of serum and then treated for 60 min at the same temperatures in the presence of 10 mM LiCl with 100 ng/ml EGF, followed by determination of
inositol phosphate formation. Background inositol phosphate formation in response to addition of EGF was determined in parallel on cells transfected with empty vector and subtracted from the individual values, with appropriate consideration of error propagation [66]. Additional experiments showed that the stimulatory effect of EGF on wild-type PLCγ2 activity was concentration-dependent with half-maximal and maximal effects at approximately 10 ng/ml and 50 ng/ml, respectively, and was almost completely blocked (~ 95 %) by the EGFR inhibitor cetuximab (*not shown*).
**Figure(s)**

A

![Diagram of PLCγ2 protein structure highlighting domains PH, 4xF, X, P, SH2n, SH2c, SH3, H, Y, C2, and regions Δ19, Δ20-22 relative to amino acid (aa) positions 0 to 1200.](image)

B

**Graph depicting inositol phosphate formation and temperature response.**

- **Inositol phosphate formation (closed symbols: cpm x 10^-3)**
  - Δ19
  - Δ20-22
  - Control
  - WT

- **Inositol phosphate formation (open symbols: cpm x 10^-3)**
  - Δ19
  - Δ20-22

- **Temperature (°C)**: 25, 27, 29, 31, 33, 35, 37, 39

C

**Western blot images showing samples labeled Control, Δ19, Δ20-22** at different temperatures (25, 27, 31, 33, 35, 37, 39°C).
Figure 2-III

E

Inositol phosphate formation (cpm x 10^3)

Time (min)

37 → 31 °C
31 → 37 °C

Δ19
Δ20-22
WT

F

1 2 3
WT Δ19 Δ20-22
-10 70 70 Time (min)
37 31 37 Temp. (°C)
Fig. 4-I
B

![Graph showing the relationship between temperature and inositol phosphate formation. The graph includes data points for WT, Δ PCI, Δ20-22, Δ PCI, Δ SA, Δ SH, and Δ SH2c, with temperature on the x-axis and inositol phosphate formation on the y-axis.](image)

Fig. 4-II
Fig. 5-1
Fig. 5-II
Fig. 6
Fig. 7
Fig. 8-II
A

![Graph showing temperature vs. inositol phosphate formation for WT and Δ19 constructs.](image)

B

![Bars comparing 31 °C and 37 °C for WT and Δ19 constructs.](image)

C

![Structure of spPHγ1 and spPHγ2 proteins with labeled residues.](image)

D

![Bar graph showing inositol phosphate formation for different constructs at 31 °C and 37 °C.](image)
Inositol phosphate formation (cpm x 10^{-3})

- **31 °C**
- **37 °C**

**Fig. 10-I**
Fig. 10-II
Fig. 10-III

Φ, Φ, ΦΦ: PtdIns(4,5)P₂ hydrolysis
SUPPLEMENTARY MATERIAL

Cool-temperature-mediated activation of phospholipase C-γ2 in the human hereditary disease PLAID

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Supplementary figures S1-S5

Fig. S1. Expression of wild-type and mutant PLCγ isozymes in Figs. 4A and 4B
Fig. S2. Expression of wild-type and mutant PLCγ isozymes in Fig. 5
Fig. S3. Expression of wild-type and mutant PLCγ isozymes in Fig. 9A
Fig. S4. Expression of wild-type and mutant PLCγ isozymes in Figs. 9B and 9D
Fig. S5. Expression of wild-type and mutant PLCγ2 isozymes and portions thereof in Fig. 10A
Fig. S6. Expression of wild-type and mutant PLCγ2 isozymes and portions thereof in Fig. 10A
Fig. S7. Expression of wild-type and mutant PLCγ2 isozymes in Fig. 11
**Supplementary Fig. S1**

**A**

![Expression of wild-type and mutant PLCγ1 and PLCγ2 isozymes in the experiment shown in Fig. 4A.](image)

**B**

![Expression of wild-type and mutant PLCγ2 isozymes in the experiment shown in Fig. 4B.](image)

**Fig. S1.** (A) Expression of wild-type and mutant PLCγ1 and PLCγ2 isozymes in the experiment shown in Fig. 4A. (B) Expression of wild-type and mutant PLCγ2 isozymes in the experiment shown in Fig. 4B. Cells from one well each were washed once with 0.2 ml of Dulbecco’s PBS and then lysed by addition of 100 µl of SDS-PAGE sample preparation buffer. The samples were subjected to SDS-PAGE and immunoblotting was performed using an antibody reactive against the c-Myc epitope on PLCγ1 or PLCγ2.
**Supplementary Fig. S2**

**A**

![Supplementary Fig. S2 A](image)

**B**

![Supplementary Fig. S2 B](image)

**C**

![Supplementary Fig. S2 C](image)

**Fig. S2.** Expression of wild-type and mutant PLCγ2 isozymes in the experiment shown in Fig. 5, **left panel, (A), center panel, (B), and right panel, (C).** Cells from one well each were analyzed by SDS-PAGE and immunoblotting using an antibody reactive against the c-Myc epitope.
Fig. S3. Expression of wild-type and mutant PLCγ2 isozymes in the experiment shown in Figs. 9A, left panel, (A) and 9A, right panel, (B). Cells from one well each were analyzed by SDS-PAGE and immunoblotting using an antibody reactive against the c-Myc epitope.
Fig. S4. Expression of wild-type and mutant PLCγ2 isozymes in the experiment shown in Figs. 9B (A) and 9D (B). Cells from one well each were analyzed by SDS-PAGE and immunoblotting using an antibody reactive against the c-Myc epitope.
**Supplementary Fig. S5**

**Fig. S5.** Expression of wild-type and mutant PLCγ2 isozymes as well as portions thereof in the experiment shown in Fig. 10A. Cells from one well each were analyzed by SDS-PAGE and immunoblotting using an antibody reactive against the c-Myc epitope.
Supplementary Fig. S6

Fig. S6. Expression of wild-type and mutant PLCγ2 isozymes as well as portions thereof in the experiment shown in Fig. 10B. Cells from one well each were analyzed by SDS-PAGE and immunoblotting using an antibody reactive against the c-Myc epitope.
**Supplementary Fig. S7**

**Fig. S7.** Expression of wild-type and mutant PLCγ2 isozymes in the experiment shown in Fig. 11. Cells from one well each were analyzed by SDS-PAGE and immunoblotting using an antibody reactive against the c-Myc epitope.