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Lowering of blood pressure after nitrate-rich vegetable consumption is abolished with the co-ingestion of thiocyanate-rich vegetables in healthy normotensive males

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ABSTRACT

A diet rich in vegetables is known to provide cardioprotection. However, it is unclear how the consumption of different vegetables might interact to influence vascular health. This study tested the hypothesis that nitrate-rich vegetable consumption would lower systolic blood pressure but that this effect would be abolished when nitrate-rich and thiocyanate-rich vegetables are co-ingested. On four separate occasions, and in a randomised cross-over design, eleven healthy males reported to the laboratory and consumed a 750 mL vegetable smoothie that was either: low in nitrate (~ 0.3 mmol) and thiocyanate (~ 5 μmol), low in nitrate and high in SCN⁻ (~ 72 μmol), high in nitrate (~ 4 mmol) and low in SCN⁻ and high in nitrate and SCN⁻. Blood pressure as well as plasma and salivary [thiocyanate], [nitrate] and [nitrite] were assessed before and 3 hours after smoothie consumption. Plasma [nitrate] and [nitrite] and salivary [nitrate] were not different after consuming the two high-nitrate smoothies, but salivary [nitrite] was higher after consuming the high-nitrate low-thiocyanate smoothie (1183 ± 625 μM) compared to the high-nitrate high-thiocyanate smoothie (941 ± 532 μM; P<0.001). Systolic blood pressure was only lowered after consuming the high-nitrate low-thiocyanate smoothie (-3 ± 5 mmHg; P<0.05). The acute consumption of vegetables high in nitrate and low in thiocyanate lowered systolic blood pressure. However, when the same dose of nitrate-rich vegetables was co-ingested with thiocyanate-rich vegetables the increase in salivary [nitrite] was smaller and systolic blood pressure was not lowered. These findings might have implications for optimising dietary guidelines aimed at improving cardiovascular health.

Key Words: Entero-salivary circulation; nitrite; nitric oxide; vascular health; nutrition
1. INTRODUCTION

Cardiovascular disease (CVD) is the leading cause of mortality globally and places a significant burden on healthcare services [1]. In an effort to attenuate CVD morbidity cost-effectively, many governments actively promote the consumption of a diet rich in fruit and vegetables [2-4] based on evidence that adhering to such diets can mitigate the risks associated with CVD morbidity [5,6]. However, emerging evidence indicates that the cardioprotective effect afforded by vegetable consumption outweighs that of fruit consumption [7], and that leafy green vegetables rich in inorganic nitrate (NO$_3^-$) might be particularly effective at improving the health of the cardiovascular system [8-10,11].

The cardiovascular benefits associated with NO$_3^-$-rich vegetable consumption has been ascribed to the stepwise reduction of NO$_3^-$ to nitrite (NO$_2^-$) and then nitric oxide (NO) [12]. Since humans have a limited capacity to directly reduce NO$_3^-$ to NO$_2^-$, with this being critically dependent on the NO$_3^-$ reducing bacteria on the tongue [13,14], a rate limiting step for the chemical reduction of NO$_3^-$ is the delivery of NO$_3^-$ to the oral cavity through the enterosalivary circulation [15]. The uptake of NO$_3^-$ into the salivary glands occurs in competition with other anions, including thiocyanate (SCN$^-$), with evidence that SCN$^-$ has a higher affinity for transport into the salivary glands compared to NO$_3^-$ [16]. It has recently been reported that cigarette smokers, who manifest higher plasma and salivary [SCN$^-$], exhibited smaller increases in salivary [NO$_3^-$] and plasma [NO$_2^-$] compared to non-smokers after ingesting the same NO$_3^-$ dose, and that blood pressure was only lowered post NO$_3^-$ supplementation in the non-smokers [17]. Therefore, increased exposure to SCN$^-$ has the potential to perturb dietary NO$_3^-$ metabolism and its beneficial effect on vascular health.
Although some green vegetables, including spinach and rocket, are a rich source of NO₃⁻ [18,19], other green vegetable varieties, including members of the Brassica family such as cabbage, cauliflower and broccoli, have been reported to increase serum [SCN⁻] [20]. Brassica vegetables are rich in glucosinolates [21,22]. During processes that damage plant cell membranes, such as mastication, glucosinolates are exposed to myrosinase which catalyses the hydrolysis of glucosinolates to SCN⁻ [23,24]. Therefore, co-ingestion of glucosinolate/SCN⁻-rich vegetables with NO₃⁻-rich vegetables has the potential to perturb dietary NO₃⁻ metabolism and the beneficial effect of NO₃⁻ on vascular health markers.

The purpose of this study was therefore to investigate the independent and combined effect of ingesting NO₃⁻-rich and SCN⁻-rich vegetables on dietary NO₃⁻ metabolism and blood pressure at doses within the recommended daily fruit and vegetable intake of 400 g·day⁻¹ [3,25]. We hypothesised that, compared to the ingestion of vegetables high in NO₃⁻ and low in SCN⁻, the increases in salivary [NO₃⁻] and [NO₂⁻] and plasma [NO₂⁻] would be blunted, and the lowering of blood pressure would be attenuated when the same dose of NO₃⁻-rich vegetables was co-ingested with SCN⁻-rich vegetables. We also hypothesised that the ingestion of vegetables low in NO₃⁻ would not alter blood pressure, irrespective of their SCN⁻ content, due to a lack of change in plasma [NO₂⁻].

2. SUBJECTS AND METHODS

2.1 Subjects characteristics

Eleven healthy, non-smoking males (mean ± SD, age 21 ± 1 yr, height 1.82 ± 0.03 m, body mass 80 ± 9 kg) were recruited from the University student community to participate in this trial. The participant number was determined a priori via a statistical power calculation.
Specifically, to detect an effect size of 1, which was based on the peak reduction in systolic blood pressure following the ingestion of NO$_3^-$-rich beetroot juice providing 4 mmol NO$_3^-$ from the study by Wylie et al. [26], with a high statistical power (0.80) and an $\alpha$ error probability of 0.05, 10 subjects were required. We recruited 11 participants to account for potential participant drop out. This calculation was conducted using G*Power (Version 3.1.9.2). All procedures employed in this study were approved by the Institutional Research Ethics Committee. Subjects gave their written informed consent to participate in this trial prior to the commencement of the study and after the experimental procedures, associated risks, and potential benefits of participation had been explained. Subjects were instructed to arrive at each laboratory testing session in a rested state after an overnight fast. Since the chemical reduction of NO$_3^-$ to NO$_2^-$ in the oral cavity is thwarted by antibacterial mouthwash [13], subjects were required to refrain from mouthwash use for the duration of the study. Each subject was also asked to avoid consumption of NO$_3^-$-rich and glucosinolate/SCN$^-$-rich foods for 48 h, and from caffeine and alcohol ingestion 12 and 24 h before each test, respectively. All subjects were instructed to maintain their habitual physical activity pattern for the duration of the study, and to avoid strenuous exercise in the 24 h preceding the testing sessions. All tests were performed at the same time of day (± 1 hour).

2.2 Experimental design

Subjects were required to report to the laboratory on five occasions over a 3-5 week period to complete the experimental testing. Since environmental (e.g. humidity, temperature, water content and exposure to sunlight) and agricultural factors can influence vegetable NO$_3^-$ and SCN$^-$ concentrations [19,22,27,28], each vegetable was obtained from the same supplier (Tesco Stores Ltd, UK) and the trial was completed within a 4-month period to limit the potential for variability in these factors.
On the first visit to the laboratory, subjects were familiarized with the blood pressure measures and venous blood and saliva collection methods described below. On the subsequent four laboratory visits, seated blood pressure was assessed, and saliva and venous blood samples were obtained, after arrival at the laboratory. Subjects then consumed a vegetable smoothie beverage prepared using vegetables either low in NO$_3^-$ and SCN$^-$ (LoN-LoT); low in NO$_3^-$, high in SCN$^-$ (LoN-HiT); high in NO$_3^-$, low in SCN$^-$ (HiN-LoT); or high in NO$_3^-$ and SCN$^-$ (HiN-HiT) along with a standardised breakfast of 54 g of porridge oats (“Oats So Simple”, Quaker Oats) prepared with 180 mL of tap water and one 20 g sachet of golden syrup (Lyle’s golden syrup). The smoothies were administered in a randomized, doubled-blind, counter-balanced, cross-over experimental design. Blood pressure was measured and venous blood and saliva were collected three hours post vegetable smoothie consumption to align with the peak plasma [NO$_2^-$] and blood pressure reduction following dietary supplementation with 4.2 mmol NO$_3^-$ [26]. Plasma and salivary [NO$_3^-$], [NO$_2^-$] and [SCN$^-$] were determined using the procedures described below. Subjects were naive to the experimental hypotheses and were informed that the aim of this study was to investigate the effects of consuming different vegetables on vascular health.

2.3 Supplementation procedures

To inform the composition of the vegetable smoothie beverages administered in this study, the NO$_3^-$ and SCN$^-$ content of a variety of vegetables was assessed prior to experimental testing. These preliminary analyses revealed that garden peas were low in both NO$_3^-$ (~ 0.8 mg/100 g) and SCN$^-$ (< 0.1 mg/100 g), spinach and rocket were high in NO$_3^-$ (~ 143.0 and 360.5 mg/100 g, respectively) and low in SCN$^-$ (both < 0.1 mg/100 g), and the inner leaves of a cabbage were high in SCN$^-$ (~ 5.0 mg/100 g) and low in NO$_3^-$ (~ 7.3 mg/100 g). Therefore,
these vegetables were selected to prepare the four vegetable smoothie beverages administered in this study. Further pilot work to assess drink palatability and the potential for side effects, including gastrointestinal (GI) discomfort, revealed that subjects were able to ingest 750 mL of each of the four vegetable smoothie beverages prepared in this study with minimal GI discomfort.

The four vegetable smoothies administered in this study were all 750 mL and comprised 250 mL tap water, 150 mL of Ribena blackcurrant cordial (Ribena, Lucozade Ribena Suntory Limited, United Kingdom) and 340 g of blended green vegetables. The 340 g of green vegetables comprised 340 g garden peas in the LoN-LoT beverage; 122 g garden peas and 218 g inner cabbage leaves in the LoN-HiT beverage; 218 g garden peas, 87 g spinach and 35 g rocket in the HiN-LoT beverage; and 218 g inner cabbage leaves, 87 g spinach and 35 g rocket in the HiN-HiT beverage. Vegetables were weighed using digital scales sensitive to 100 mg (Ohaus Valor 3000 Xtreme, Ohaus Corporation, USA) and blended on the morning of testing. Each smoothie was then refrigerated at 4°C for > 15 min prior to consumption. Vegetables were blended raw to preserve nutritional content, since conventional cooking methods reduce the intake of glucosinolates/SCN⁻ [22] and NO₃⁻ [29]. A 30 mL aliquot was obtained from each of the four vegetable smoothies from which subsequent 1.5 mL aliquots were collected and analyzed to determine the NO₃⁻ and SCN⁻ content of the vegetable smoothie beverages (see below for further details). These analyses were completed at the start and end of the study to gauge any changes in the vegetable NO₃⁻ and SCN⁻ content across the study. The NO₃⁻ and SCN⁻ content of the four vegetable smoothie beverages at the start and end of the study are presented in table 1, with the mean of these two samples used to indicate the mean NO₃⁻ and SCN⁻ contents of the beverages across the duration of the study.
2.4 Measurements

2.4.1 Blood pressure

Subjects were required to rest seated for 10 min in an isolated temperature-controlled room. Thereafter, blood pressure of the brachial artery was measured whilst the subject was seated using an automated sphygmomanometer (Dinamap Pro, GE Medical Systems, Tampa, USA). Five measurements were taken and the mean of the five measurements was used for analysis.

2.4.2 Blood and saliva collection

Venous blood samples were drawn into 6 mL lithium-heparin tubes (Sarstedt, Leicester, UK). Samples were centrifuged at 4,000 rpm and 4°C for 10 min, within 1 min of collection. Plasma was subsequently extracted and immediately frozen at -80°C for later analysis of [NO$_3^-$], [NO$_2^-$] and [SCN$^-$. Unstimulated saliva samples (~ 3 mL) were collected into 30 mL universal containers and 1 mL aliquots were frozen at -80°C for later analysis of [NO$_3^-$], [NO$_2^-$] and [SCN$^-$.]

2.5 Data analysis procedures

2.5.1 Plasma and salivary [NO$_3^-$] and [NO$_2^-$] determination

All glassware, utensils, and surfaces were rinsed with deionized water to remove residual NO intermediates prior to [NO$_3^-$] and [NO$_2^-$] analysis. Plasma samples were deproteinized using zinc sulfate (ZnSO$_4$/sodium hydroxide (NaOH) precipitation prior to determination of [NO$_3^-$]. Firstly, 500 µL of 0.18 N NaOH was added to 100 µL of sample followed by a 5 min incubation at room temperature. Subsequently, samples were treated with 300 µL of aqueous ZnSO$_4$ (5% w/v) and vortexed for 30 seconds before undergoing an additional 10 min incubation period at room temperature. Samples were then centrifuged at 4,000 rpm for 5 min, and the supernatant was removed for subsequent analysis. The [NO$_3^-$] of the
deproteinized plasma sample was determined by its reduction to NO in the presence of 0.8% (w/v) vanadium chloride (VCl₃) in 1 M HCl within an air-tight purging vessel. Plasma samples were introduced to the vessel via 50 μL injections into the septum at the top of the vessel. The spectral emission of electronically excited nitrogen dioxide, derived from the reaction of NO with ozone, was detected by a thermoelectrically cooled, red-sensitive photomultiplier tube housed in a Sievers gas-phase chemiluminescence nitric oxide analyzer (Sievers NOA 280i, Analytix Ltd, Durham, UK). The [NO₃⁻] was determined by plotting signal (mV) area against a calibration plot of sodium nitrate standards. The [NO₂⁻] of the undiluted (non-deproteinized) plasma was determined by its reduction to NO in the presence of glacial acetic acid and aqueous sodium iodide (4% w/v). Sodium nitrite standards were used. Injections of 100 μL of plasma were used for [NO₂⁻] determination. After thawing at room temperature, saliva samples were centrifuged for 10 min at 14000 rpm and the supernatant was removed for subsequent analysis. The supernatant was diluted 100-fold with deionized water and [NO₃⁻] and [NO₂⁻] were determined from 50 μL injections, using the same reagents describe above for the plasma analyses.

2.5.2 Plasma and salivary [SCN⁻] determination
Plasma and salivary [SCN⁻] were measured in duplicate using the procedures described by Tsuge et al. [30]. Briefly, 300 μL of plasma was treated with 200 μL of trichloroacetic acid (25% w/v) and centrifuged for 10 min at 14000 rpm. The supernatant was then removed for subsequent analysis. 50 μL of sample, 15 μL of 1 M of ice-cold potassium hydrogen phosphate solution (pH 5.5), 5 μL of 5M NaOH solution, and 10 μL of 6.25 mg/ml chloramine T solution were added to a 96-well microtiter plate (Sterilin Ltd., Caerphilly, UK) and incubated on ice for 2 min. Subsequently, 120 μL of cyanoline blue solution (0.27% w/v) was added to the microtiter plate and the plate was incubated for 20 min at room
Following incubation, sample absorbance was measured at 620 nm using a microplate reader (EnSpire 2300, Perkin Elmer, Hamburg, Germany). Thawed saliva samples were centrifuged for 10 min at 14000 rpm and the supernatant was removed for subsequent analysis. Samples were diluted 25 fold, and 40 μL of the diluted sample was added to a 96-well microtiter plate in addition to 20 μL of 1 M potassium phosphate buffer solution and 20 μL of 6.25 mg/ml choramine T solution, and incubated on ice for 2 min. Cyanoline blue solution (140 μL 0.27 % w/v in pyridine-water, 1:5, v/v) was added to the microtiter plate followed by a 20 min incubation at room temperature. Following incubation, sample absorbance was measured at 620 nm using a microplate reader.

2.5.3 Vegetable smoothie beverage [NO₃⁻] and [SCN⁻] determination

Prior to analysis of the vegetable smoothie beverage NO₃⁻ content, a 1.5 mL aliquot of each vegetable smoothie beverage was transferred to a heat-resistant microcentrifuge tube. Subsequently, samples were heated at 130 ºC for 60 min using a heat plate (Grant QBD2, Cambridge, UK) to disintegrate the cell membranes for release of intracellular NO₃⁻. Samples were then centrifuged at 12,000 g and 4 ºC for 8 min and the supernatant was removed for subsequent analysis. The supernatant was diluted 100-fold with deionized water and [NO₃⁻] was determined from 50 μL injections using the same reagents and procedures described above for plasma [NO₃⁻] analysis.

For determination of the vegetable smoothie beverage SCN⁻ content, a 1.5 mL aliquot of each vegetable smoothie beverage was transferred to a standard microcentrifuge tube. These samples were not heated, as it was necessary to avoid the denaturation of the enzyme, myrosinase, which hydrolyzes glucosinolates to SCN⁻ [22]. Vegetable smoothie samples were centrifuged at 12,000 g and 4 ºC for 10 min and the supernatant was removed for
subsequent analysis. The [SCN\(^{-}\)] of the supernatant was determined using the same reagents and procedures described above for salivary [SCN\(^{-}\)] analysis.

2.6 Statistical analysis

A two-way (supplement × time) repeated-measures ANOVA was employed to assess pre-post supplement changes in blood pressure variables and plasma and salivary [NO\(_3^{-}\)], [NO\(_2^{-}\)] and [SCN\(^{-}\)] for the four vegetable smoothie beverages administered in this study (LoN-LoT, LoN-HiT, HiN-LoT and HiN-HiT). Where the ANOVA revealed significant differences, Fishers Least Significant Difference tests were employed to determine the origin of such effects. Pearson’s product moment correlation coefficient was used to assess the relationship between changes in variables across conditions. All data are presented as mean ± SD unless otherwise indicated. Statistical significance was accepted when \(P<0.05\). All statistical analyses were conducted using IBM SPSS Statistics version 23.

3. RESULTS

Participants self-reported that their diet and exercise patterns were consistent for the 48 hours preceding each laboratory visit.

3.1 Plasma and salivary [SCN\(^{-}\)]

The plasma and salivary [SCN\(^{-}\)] across the four experimental conditions are illustrated in figure 1. There was a significant supplement × time interaction effect for plasma and salivary [SCN\(^{-}\)] (both \(P<0.001\)). Compared to the pre-supplementation values, plasma [SCN\(^{-}\)] was higher in the LoN-HiT (12 ± 4 vs. 20 ± 7 μM) and HiN-HiT (11 ± 4 vs. 20 ± 6 μM) conditions (\(P<0.01\)), but not different in the LoN-LoT (16 ± 6 vs. 15 ± 8 μM) and HiN-LoT (16 ± 5 vs. 17 ± 7 μM) conditions (\(P>0.05\); Figure 1). Likewise, compared to the pre-
supplementation values, salivary [SCN\(-\)] was higher post smoothie consumption in the LoN-HiT (1061 ± 377 vs. 1463 ± 378 µM) and HiN-HiT (976 ± 519 vs. 1286 ± 393 µM) conditions (\(P<0.01\)), but not different in the LoN-LoT (1219 ± 495 vs. 1176 ± 575 µM) and lower in the HiN-LoT (1283 ± 485 vs. 922 ± 368 µM) conditions (\(P>0.05\); Figure 1).

3.2 Plasma and salivary \([\text{NO}_3^-]\) and \([\text{NO}_2^-]\)

The plasma and salivary \([\text{NO}_3^-]\) and \([\text{NO}_2^-]\) across the four experimental conditions are illustrated in figure 2. There were significant supplement × time interaction effects for plasma and salivary \([\text{NO}_3^-]\) and \([\text{NO}_2^-]\) (\(P<0.001\) for all comparisons). The post-supplementation plasma \([\text{NO}_3^-]\) was higher than the pre-supplementation values in the HiN-LoT (44 ± 24 vs. 276 ± 62 µM) and HiN-HiT (42 ± 13 vs. 257 ± 54 µM) conditions (\(P<0.001\)), but not the LoN-LoT (44 ± 30 vs. 41 ± 20 µM) and LoN-HiT (42 ± 17 vs. 41 ± 14 µM) conditions (\(P>0.05\); Figure 2). Moreover, the post-supplementation plasma \([\text{NO}_3^-]\) values were higher in the HiN-LoT and HiN-HiT conditions compared to both the LoN-LoT and LoN-HiT conditions (\(P<0.001\)), but were not different between the HiN-LoT and HiN-HiT conditions (\(P>0.05\)). Salivary \([\text{NO}_3^-]\) was higher post-supplementation compared to pre-supplementation in the HiN-LoT (513 ± 705 vs. 3114 ± 2110 µM) and HiN-HiT (382 ± 525 vs. 3362 ± 2232 µM) conditions (\(P<0.001\)), but not the LoN-LoT (445 ± 438 vs. 403 ± 387 µM) and LoN-HiT (428 ± 375 vs. 315 ± 252 µM) conditions (\(P>0.05\); Figure 2). The post-supplementation salivary \([\text{NO}_3^-]\) values were also higher in the HiN-LoT and HiN-HiT conditions compared to both the LoN-LoT and LoN-HiT conditions (\(P<0.001\)), but were not different between the HiN-LoT and HiN-HiT conditions (\(P>0.05\)). The change in the pre-to-post-supplementation plasma and salivary \([\text{NO}_3^-]\) were not different between the HiN-LoT and HiN-HiT conditions (\(P>0.05\)).
Post-supplementation plasma $[\text{NO}_2^-]$ was higher than the pre-supplementation values in the HiN-LoT (121 ± 32 vs. 327 ± 110 nM) and HiN-HiT (137 ± 59 vs. 336 ± 108 nM) conditions ($P<0.001$), but not the LoN-LoT (123 ± 45 vs. 115 ± 40 nM) and LoN-HiT (117 ± 37 vs. 123 ± 37 nM) conditions ($P>0.05$; Figure 2). The post-supplementation plasma $[\text{NO}_2^-]$ values were higher in the HiN-LoT and HiN-HiT conditions compared to both the LoN-LoT and LoN-HiT conditions ($P<0.001$), but were not different between the HiN-LoT and HiN-HiT conditions ($P>0.05$). Salivary $[\text{NO}_2^-]$ was higher post-supplementation compared to pre-supplementation in the HiN-LoT (233 ± 127 vs. 1183 ± 625 µM) and HiN-HiT (171 ± 107 vs. 941 ± 532 µM) conditions ($P<0.001$), but not the LoN-LoT (289 ± 220 vs. 192 ± 118 µM) and LoN-HiT (283 ± 249 vs. 177 ± 123 µM) conditions ($P>0.05$; Figure 2). The post-supplementation salivary $[\text{NO}_2^-]$ values were also higher in the HiN-LoT and HiN-HiT conditions compared to both the LoN-LoT and LoN-HiT conditions ($P<0.001$). However, salivary $[\text{NO}_2^-]$ was higher post-supplementation in the HiN-LoT condition compared to the HiN-HiT condition ($P<0.05$; Figure 2). The change in the pre-to-post-supplementation plasma $[\text{NO}_2^-]$ was not different between the HiN-LoT and HiN-HiT conditions ($P>0.05$), but there was a trend ($P=0.09$) for a greater increase in pre-to-post-supplementation salivary $[\text{NO}_2^-]$ in the HiN-LoT condition compared to the HiN-HiT condition.

### 3.3 Blood pressure

The systolic, diastolic and mean arterial blood pressures across the four experimental conditions are illustrated in figure 3. Systolic blood pressure was not different pre- and post-supplementation in the LoN-LoT (119 ± 7 vs. 119 ± 8 mmHg), LoN-HiT (118 ± 5 vs. 117 ± 8 mmHg) and HiN-HiT (119 ± 6 vs. 119 ± 7 mmHg) conditions ($P>0.05$), but was lower post-supplementation in the HiN-LoT condition (118 ± 6 vs. 115 ± 5 mmHg; $P<0.05$; Figure 3). Diastolic blood pressure was not different pre- and post-supplementation in the LoN-LoT (66
± 5 vs. 64 ± 5 mmHg), LoN-HiT (65 ± 5 vs. 63 ± 5 mmHg), HiN-LoT (63 ± 5 vs. 64 ± 5 mmHg) and HiN-HiT (64 ± 4 vs. 64 ± 6 mmHg) conditions (P>0.05). Similarly, mean arterial pressure was not different pre- and post-supplementation in the LoN-LoT (83 ± 4 vs. 82 ± 5 mmHg), LoN-HiT (83 ± 5 vs. 81 ± 5 mmHg), HiN-LoT (82 ± 4 vs. 81 ± 4 mmHg) and HiN-HiT (83 ± 3 vs. 83 ± 6 mmHg) conditions (P>0.05). The lowering in systolic blood pressure in the HiN-LoT condition was not correlated with the increase in salivary [NO$_2^-$] (r = 0.08; P>0.05). Moreover, there was no correlation between the differences in systolic blood pressure and salivary [NO$_2^-$] post smoothie ingestion between the HiN-LoT condition and the LoN-LoT (r = -0.02; P>0.05) and HiN-HiT (r = 0.06; P>0.05) conditions.

4. DISCUSSION

The important novel finding from this study was that co-ingestion of NO$_3^-$-rich and SCN$^-$-rich vegetables blunted the increase in salivary [NO$_2^-$] and the lowering of systolic blood pressure compared to the same portion of NO$_3^-$-rich vegetables ingested without SCN$^-$-rich vegetables. The ingestion of vegetables low in both NO$_3^-$ and SCN$, and low in NO$_3^-$ but high in SCN$, did not impact plasma and salivary [NO$_3^-$] and [NO$_2^-$] or blood pressure responses. These results may have a potentially important public health message by suggesting that consuming 340 g of green vegetables, a dose within the recommended daily intake of fruit and vegetables [3,24], evokes disparate effects on blood pressure dependent on the composition of the vegetables consumed. Specifically, it appears that blood pressure is lowered following the acute consumption of NO$_3^-$-rich vegetables without co-ingestion of SCN$^-$-rich vegetables, but not after consumption of vegetables rich in SCN$^-$ and low in NO$_3^-$ or when NO$_3^-$-rich and SCN$^-$-rich vegetables are co-ingested. Our findings may have implications for finessing dietary guidelines aimed at enhancing cardiovascular health, particularly since current
nutritional guidelines do not differentiate between low- and high-NO$_3^-$ vegetable consumption [3,4].

When the cell membranes of glucosinolate-rich plants are damaged, as occurs during mastication and during the blending procedures used to prepare the vegetable smoothie beverages administered in this study, glucosinolates are presented to myrosinase and subsequently hydrolysed to form SCN$^-$ [23,24]. Therefore, our observation of increased salivary and plasma [SCN$^-$] after consuming a glucosinolate-rich Brassica vegetable beverage in the LoN-HiT and HiN-HiT conditions is likely a function of myrosinase-catalyzed glucosinolate hydrolysis, and is consistent with previous reports of increased serum [SCN$^-$] after the consumption of Brassica vegetables [20]. In addition, and also consistent with previous studies, ingestion of NO$_3^-$-rich vegetables in the HiN-LoT and HiN-HiT conditions increased salivary and plasma [NO$_3^-$] and [NO$_2^-$] [8,13,14,17,26].

Salivary [SCN$^-$] was lowered post supplementation compared to pre supplementation in the HiN-LoT condition consistent with previous observations [17] and with the notion that NO$_3^-$ and SCN$^-$ compete for a common salivary transporter [16,17]. However, in contrast with our hypothesis, salivary [NO$_3^-$] increased by a similar magnitude in the HiN-LoT and HiN-HiT conditions despite increased salivary and plasma [SCN$^-$] in the latter compared to the former. This finding also conflicts with our previous observation that the increase in salivary [NO$_3^-$] after NO$_3^-$ ingestion was lower in smokers, who exhibited elevated plasma and salivary [SCN$^-$], compared to non-smokers [17]. These conflicting findings are likely attributable to greater differences in salivary (+ 1164 µM) and plasma (+ 22 µM) [SCN$^-$] between the smokers and non-smokers in the control condition of our previous study [17] compared to the mean change in salivary (+ 356 µM) and plasma (+ 9 µM) [SCN$^-$] after consuming the two HiT beverages.
administered in the current study. Therefore, the findings in the present study suggest that increasing dietary SCN’ intake, via the consumption of glucosinolate-rich vegetables, is unlikely to perturb salivary NO$_3^-$ uptake at doses within the recommended daily fruit and vegetable consumption guidelines [3,25]. However, whilst the increase in salivary [NO$_3^-$] was similar in the HiN-LoT and HiN-HiT conditions, the increase in salivary [NO$_2^-$] above baseline tended ($P=0.09$) to be higher in the HiN-LoT condition compared to the HiN-HiT condition and the post-supplementation salivary [NO$_2^-$] was higher in the HiN-LoT condition compared to the HiN-HIT condition. This suggests that, at the lower exposure rates applied in the current study, SCN’ has the potential to impede the reduction of NO$_3^-$ to NO$_2^-$ in the oral cavity, or to catalyze NO$_2^-$ degradation, rather than interfering with salivary NO$_3^-$ uptake, as was observed at higher SCN’ exposure rates in our recent smoking study [17]. The lower salivary [NO$_2^-$] in the HiN-HiT condition compared to the HiN-LoT condition might be linked to SCN’-mediated activation of myeloperoxidase (MPO), which oxidizes NO$_2^-$ to nitrogen dioxide (NO$_2$) [31]. In addition, MPO catalyses the oxidation of chloride (Cl$^-$) to hypochlorite (OCl$^-$) [32] which can react with SCN’ to form hypothiocyanite (OSCN$^-$) [33], a potent oral antimicrobial agent [34]. OSCN$^-$ can also be produced from SCN’ through the activity of salivary peroxidase (SPO) [35]. Therefore, the increase in salivary [SCN$^-$] in the HiN-HiT condition might have activated MPO and SPO resulting in increased synthesis of the antimicrobial species, OSCN$^-$ [34], and a corresponding blunting of the rate of NO$_3^-$ reduction to NO$_2^-$ by the oral NO$_3^-$ reducing bacteria [13,14], compared to the HiN-LoT condition.

Systolic blood pressure was lowered in the HiN-LoT condition but not the LoN-LoT, LoN-HiT and HiN-HiT conditions in this study, consistent with our hypotheses. This 3 mmHg reduction in systolic blood pressure in the HiN-LoT condition is consistent with the lowering of systolic blood pressure after the consumption of NO$_3^-$-rich spinach [36,37], rocket [27],
beetroot [11] or a mixture of NO₃⁻-rich vegetables [8]. The magnitude by which blood pressure is lowered in these studies has potential clinical relevance having been associated with a lower incidence of future cardiovascular events [38]. It should also be acknowledged that systolic blood pressure has been reported to be lowered to a greater extent after ingesting 8 mmol compared to 4 mmol of NO₃⁻ (the same dose as administered in the current study in the NO₃⁻-rich smoothies) when administered in the form of NO₃⁻-rich beetroot juice [26]. Therefore, while it is likely that we could have observed a greater fall in systolic blood pressure with a greater NO₃⁻ dose, the requirements to combine different combinations of NO₃⁻-rich, SCN⁻-rich, NO₃⁻-low and SCN⁻-low vegetables, and to not exceed the recommended daily fruit and vegetable intake of 400 g·day⁻¹ [3,25] restricted our ability to administer a higher NO₃⁻ dose in the current study.

The lowering of blood pressure after the consumption of NO₃⁻-rich vegetables has been attributed to increased circulating plasma [NO₂⁻] [11,13,14,17,26,39,40], which can impact vascular function directly [41] or through the O₂-independent reduction of NO₂⁻ to NO production [42]. These changes in blood pressure after NO₃⁻ ingestion might be linked to increased classical NO-cyclic guanosine monophosphate (cGMP) signalling [39] or changes in renal physiology [43]. Therefore, the lack of change in systolic blood pressure in the LoN-LoT and LoN-HiT conditions is likely a function of a lack of increase in plasma [NO₂⁻] in these conditions. However, plasma [NO₂⁻] was increased by a similar magnitude in the HiN-LoT and HiN-HiT conditions, but blood pressure was only lowered in the former. Although plasma [NO₂⁻] was increased to a similar extent in the HiN-LoT and HiN-HiT conditions, the increase in salivary [NO₂⁻] was greater in the HiN-LoT condition. The increased salivary [NO₂⁻] in the HiN-LoT condition would have been expected in increase NO₂⁻ delivery to the stomach, where the acidic pH facilitates the chemical reduction of NO₂⁻ to NO and a number
of reactive nitrogen intermediates (RNI) including S-nitrosothiols (RSNO) [44]. It has recently been reported that the antihypertensive effect of oral NO$_2^-$ administration is closely associated with plasma [RSNO], but can be dissociated from plasma [NO$_2^-$] [45,46]. Therefore, whilst plasma [NO$_2^-$] was not different between the HiN-LoT and HiN-HiT conditions, it is possible that the total circulating [RSNO] pool was greater in the HiN-LoT condition leading to a lower blood pressure in this condition compared to the HiN-HiT condition.

There is evidence that SCN$^-$ can lower the acidity of the gastric juice [47]. Combined with a potential lowering in gastric NO$_2^-$ delivery, this might have contributed to a lowered gastric RSNO formation and passage into the circulation, and explained the lack of change in systolic blood pressure in the HiN-HiT condition and the lowering of systolic blood pressure in the HiN-LoT condition. However, since SCN$^-$ has been reported to aid NO synthesis in the stomach [44,48], which through subsequent oxidation and autoxidation reactions might lead to the synthesis of RSNO [44,49], and since RSNO was not determined in the current study, further research is required to assess how different combinations of vegetables with varying NO$_3^-$ and SCN$^-$ contents influences circulating [RSNO] and blood pressure. Moreover, the lowering of systolic blood pressure and the increase in salivary [NO$_2^-$] in the HiN-LoT condition, and the differences in systolic blood pressure and salivary [NO$_2^-$] post smoothie ingestion between the HiN-LoT condition and the LoN-LoT and HiN-HiT conditions were not correlated in the current study. Accordingly, the link between the greater reduction of blood pressure and increase in salivary [NO$_2^-$], and by extension stomach NO$_2^-$ delivery, in the HiN-LoT condition compared to the HiN-HiT is unclear and in need of further research. It should also be acknowledged that, although the NO$_3^-$ and SCN$^-$ content of the smoothies administered in the current study was quantified and controlled, we cannot exclude the
possibility that other nutrients differed between the smoothies and influenced systolic blood pressure either independently or synergistically with NO$\text{$_3^-$}$ or NO$\text{$_2^-$}$. For example, ascorbic acid and polyphenols [50-52] can aid the reduction of NO$\text{$_2^-$}$ to NO in the stomach which might have influenced our systolic blood pressure results. Further research is required to resolve the underlying mechanisms for a lowering of systolic blood pressure in the HiN-LoT condition but not the HiN-HiT condition.

Although blood pressure was not improved in the LoN-HiT and HiN-HiT conditions in this study, glucosinolate-rich vegetables possess anti-cancer properties [53, 54] and their consumption may have other benefits, including a reduction in death from atherosclerotic vascular disease [55]. As such, consumption of glucosinolate-rich vegetables should certainly not be avoided, but further research is required to assess the longer-term and wider health implications of consuming different combinations of vegetables. Moreover, since some cooking methods would denature myrosinase [22], and therefore the potential for SCN$^-$ ingestion, it is unclear whether these findings can be reproduced if glucosinolate-rich vegetables are cooked prior to consumption.

In conclusion, the acute consumption of vegetables high in NO$\text{$_3^-$}$ and low in SCN$^-$, lowered systolic blood pressure in association with increased plasma and salivary [NO$\text{$_3^-$}$] and [NO$\text{$_2^-$}$]. However, when the same portion of high NO$\text{$_3^-$}$ vegetables was co-ingested with high SCN$^-$ vegetables the lowering of blood pressure was abolished and the increase in salivary [NO$\text{$_2^-$}$] was attenuated. Plasma and salivary [NO$\text{$_3^-$}$] and [NO$\text{$_2^-$}$] were not increased and systolic blood pressure was not lowered after consuming vegetables low in NO$\text{$_3^-$}$ and high SCN$^-$ or low in both NO$\text{$_3^-$}$ and SCN$^-$. These results suggest that green vegetables are not uniform in their potential to improve blood pressure and that NO$\text{$_3^-$}$-rich vegetables can lower blood pressure
when consumed with vegetables low, but not high, in SCN. Therefore, it appears that the potential of green vegetables to improve blood pressure, at portion sizes (340 g) within the recommended daily fruit and vegetable intake guidelines, is dependent on the nutritional composition of the vegetables consumed. These findings might have implications for improving public health by helping to refine nutritional guidelines aimed at improving cardiovascular health and lowering the risk of cardiovascular disease morbidity.
REFERENCES


**Figure legends**

*Figure 1.* Plasma (upper panel) and salivary (lower panel) thiocyanate concentration ([SCN⁻]) in the low nitrate low thiocyanate (LoN-LoT), low nitrate high thiocyanate (LoN-HiT), high nitrate low thiocyanate (HiN-LoT) and high nitrate high thiocyanate (HiN-HiT) vegetable smoothie conditions. Group mean (± SEM) responses prior to vegetable smoothie ingestion are shown as open bars with responses following vegetable smoothie ingestion shown as filled bars. Individual responses to the vegetable smoothies are shown for each experimental condition as solid grey lines. * indicates that the post supplementation value was significantly different to the pre supplementation value within the same experimental condition (P<0.01).

*Figure 2.* Plasma (panel A) and salivary (panel B) nitrate concentration ([NO₃⁻]) and plasma (panel C) and salivary (panel D) nitrite concentration ([NO₂⁻]) in the low nitrate low thiocyanate (LoN-LoT), low nitrate high thiocyanate (LoN-HiT), high nitrate low thiocyanate (HiN-LoT) and high nitrate high thiocyanate (HiN-HiT) vegetable smoothie conditions. Group mean (± SEM) responses prior to vegetable smoothie ingestion are shown as open bars with responses following vegetable smoothie ingestion shown as filled bars. Individual responses to the vegetable smoothies are shown for each experimental condition as solid grey lines. * indicates that the post supplementation value was significantly different to the pre supplementation value within the same experimental condition (P<0.01). # indicates that the post supplementation value was significantly different to the post supplementation value in the HiN-LoT condition (P<0.05).
Figure 3. Systolic (panel A), diastolic (panel B) and mean arterial (panel C) blood pressures in the low nitrate low thiocyanate (LoN-LoT), low nitrate high thiocyanate (LoN-HiT), high nitrate low thiocyanate (HiN-LoT) and high nitrate high thiocyanate (HiN-HiT) vegetable smoothie conditions. Group mean (± SEM) responses prior to vegetable smoothie ingestion are shown as open bars with responses following vegetable smoothie ingestion shown as filled bars. Individual responses to the vegetable smoothies are shown for each experimental condition as solid grey lines. * indicates that the post supplementation value was significantly different to the pre supplementation value within the same experimental condition ($P<0.05$). Please note that in panel A two participants had the same systolic blood pressure values before and after the ingestion of the HiN-LoT beverage so there are only 10 visible individual response lines for this condition.