Small-scale skeletal muscle constructs for in-vitro musculoskeletal junction preclinical testbed [Abstract]

This item was submitted to Loughborough University's Institutional Repository by the/an author.

Citation: WRAGG, N.M. ... ET AL., 2013. Small-scale skeletal muscle constructs for in-vitro musculoskeletal junction preclinical testbed. European Cells and Materials (ECM), 26 (Suppl. 1), pp. 93.

Additional Information:
- This is a conference abstract presented at the Tissue and Cell Engineering Society (TCES) meeting 23rd - 25th July 2013, Cardiff Institute of Tissue Engineering and Repair (CITER), Cardiff University. This is an Open Access Article. It is published by European Cells & Materials Ltd under the Creative Commons Attribution-ShareAlike 4.0 International (CC BY-SA 4.0) licence. Full details of this licence are available at: http://creativecommons.org/licenses/by-sa/4.0/

Metadata Record: https://dspace.lboro.ac.uk/2134/37761

Version: Published

Publisher: © The authors. Published by European Cells & Materials Ltd

Rights: This work is made available according to the conditions of the Creative Commons Attribution-ShareAlike 4.0 International (CC BY-SA 4.0) licence. Full details of this licence are available at: http://creativecommons.org/licenses/by-sa/4.0/

Please cite the published version.
Small-scale skeletal muscle constructs for *in-vitro* musculoskeletal junction pre-clinical testbed.

NM Wragg¹, DJ Player¹, Y Liu², MP Lewis¹

¹Musculoskeletal Biology Research Group, School of Sport, Exercise and Health Sciences, Loughborough University
²Healthcare Engineering Group, School of Mechanical and Manufacturing Engineering, Loughborough University

**INTRODUCTION:** Currently, musculoskeletal tissue damage is surgically repaired with the aim of restoring functionality over form. Any resulting tissue damage reduces utility and can increase the likelihood of further local injury. Therefore, more effective repair and regeneration techniques are needed.

By creating an in vitro musculoskeletal junction that closely replicates the form and function of the in vivo system, novel materials and chemical products could be tested for viability in a more representative manner.

In order to engineer a repeatable system capable of scaling-up, a small scale model of bone and muscle must be optimised.

**METHODS:** 0.75mL 3D type-I rat-tail collagen (2.20mg/mL) neutralised constructs were seeded with C2C12 murine myoblast cells at 4x10⁶ cells/mL as previously described (Sharples et al. 2012) and set in a gelling area of 11mm x 14mm at 37°C, 5% CO₂. The constructs were tethered at either end by bespoke polythene mesh floatation bars to create longitudinal lines of isometric tension (Fig 1). Constructs were placed in 20% FBS high glucose DMEM for 4 days and then cultured in 2% horse serum high glucose DMEM to induce differentiation.

**RESULTS & DISCUSSION:** Immunohistochemical staining for the intermediate filament protein Desmin showed the capacity for alignment and differentiation of C2C12 myoblasts within the collagen system at this scale (Fig 1). This replicates at a molecular level what is seen in larger constructs (Sharples et al. 2012).

**CONCLUSIONS:** Preliminary indications show that a small-scale tissue engineered muscle suitable for a preclinical test-bed is viable from a phenotypic standpoint. Myotubes are evident and aligned however genotypic considerations and co-cultured limitations with osteoblasts are yet to be taken into account.


**ACKNOWLEDGEMENTS:** With thanks to the EPSRC as the funding body.